Degradation of iohexol by UV irradiation: kinetics, pathways and iodinated trihalomethanes formation during post-chlorination

Yan-Zheng Liu^a, Xiao-Chang Wang^a, Fu-Xiang Tian^{b,*}, Hai-Bo Xing^b, Rong Chen^a, Ying Shen^a

^aCenter for Urban Alternative Water Resources Development, School of Environmental and Municipal Engineering, Xi'an University of Architecture and Technology, Xi'an, 710055, China, Tel. +86 29 82202121, Fax +86 29 82202729, email: liuyanzheng1@126.com (Y.-Z. Liu), xcwang@xauat.edu.cn (X.-C. Wang), chenrong@xauat.edu.cn (R. Chen), 452004434@qq.com (Y. Shen)

^bSchool of Chemical and Environmental Engineering, Shanghai Institute of Technology, Shanghai, 201418, China, Tel. +86 29 82202121, Fax +86 29 82202729, email: 13916841275@163.com (F.-X. Tian), michaelyanzi@163.com (H.-B. Xing)

Received 6 November 2016; Accepted 29 May 2017

ABSTRACT

The photodegradation of iohexol by UV irradiation and the formation of iodinated trihalomethanes (I-THMs) during subsequent chlorination were investigated in this study. Iohexol can be photodegraded by UV with pseudo-first order kinetics and the quantum yield was determined to be 0.0385 mol einstein⁻¹. The degradation rate of iohexol was linear along with the increase of UV intensity while effects of solution pH and Br[−] were negligible. The main photodegradation intermediates including deiodinated and hydroxylated products were identified by UPLC-ESI-MS. Then the UV photodestruction pathways of iohexol were proposed. The I-THMs formation in UV-chlorination of iohexol was evaluated in terms of UV fluence, pH, Br[−] concentration and raw water matrix. The increase of UV fluence and Br[−] concentration significantly enhanced the I-THMs formation and iodine substitution factor (ISF) while effect of pH was negligible. Raw freshwater matrix can appreciably influence the photodegradation rate of iohexol and I-THMs formation due to the presence of natural organic matter and Br[−]. It was found that the UV removal of iohexol in real waters was depressed while I-THMs formation and ISF values were greatly enhanced. The results of the current study led to some practical concerns on toxic I-THMs in the UV application of iohexol-containing raw waters.

Keywords: Iohexol; UV irradiation; Iodinated trihalomethanes (I-THMs); Chlorination

1. Introduction

A great variety of pharmaceuticals and personal care products (PPCPs) have been widely used and therefore are frequently detected in aquatic environment as emerging contaminants [1–3]. As representative PPCPs that catch scholars' attention, iodinated X-ray contrast media (ICM) have been detected in effluents of wastewater treatment plants and surface waters at elevated concentrations [4–8]. A study in U.S. drinking water sources found that ICM were detected at several ng L^{-1} to 2.7 µg L^{-1} (iopamidol) and 120 ng L⁻¹ (iohexol) [9]. The occurrence of ICM in German was also reported at μ g L⁻¹ level and the maximum reached as high as 100 μ g L⁻¹ [8]. Iopamidol and iohexol are the most frequently detected ICM in waters reported to date [5,9]. ICM are not toxic in themselves, but their presence can lead to the formation of iodinated disinfection by-products (I-DBPs) [9]. Besides, when chlorinated, ICM, especially iohexol, generated an enhanced level of CHO cell cytotoxicity and genotoxicity [10].

I-DBPs, including iodo-acids and iodo-trihalomethanes (I-THMs), are believed to be much more toxic than their chlorinated and brominated analogues [11,12]. Besides, the occurrence of I-THMs (especially iodoform) in drinking water may cause serious taste and odor problems [13].

^{*}Corresponding author.

^{1944-3994 / 1944-3986 © 2017} Desalination Publications. All rights reserved.

The concentrations of I-THMs were reported at µg L⁻¹ level in chloraminated and chlorinated drinking waters in the United States and Canada [8]. Therefore, many studies focused on the formation of I-DBPs have been developed, making it a hot point and academic front in water treatment area [9,14–18]. The iodine sources contributing to the I-DBPs formation incorporate iodide (I⁻) [13, 19–21], iodinebased disinfectants [22], and ICM [9,15,16]. I- can be oxidized to HOI by oxidant and then I-DBPs were formed in the presence of natural organic matter (NOM) [13,19-21]. Iodoform can generate during disinfection of source waters by iodine tincture or iodine tablets [22]. ICM have been evidenced to be effective organic iodine source to form I-DBPs [9,16]. Moreover, the nontoxic ICM (iopamidol) could release I- from UV photodegradation and transform to harmful I-DBPs in sequential oxidation processes [15]. Research on the transformation of potentially toxic ICM to I-DBPs by different treatment is of practical significance to water quality regulation.

UV irradiation has many advantages over chemical disinfection, such as producing no common halogenated DBPs, decreasing the treatment cost and inactivation a wide range of waterborne pathogens [23,24]. Lots of UV degradation studies have been reported and results showed that higher UV intensity and lower initial contaminant concentration led to greater degradation rate while the pH effect was negligible [3,15,25–27]. But the major deficiency of UV disinfection is that it can not maintain a disinfectant residual [24,28], thus the chlorine-based oxidants are often added to resupply residual disinfection ability after UV process.

The extremely stable structure of ICM makes these triiodinated compounds persistent to conventional water treatment processes [5–7], even ozonation can only remove 35–55% of non-ionic ICM [4]. Due to the strong oxidative capacity of hydroxyl radicals, advanced treatment processes have proved successful in degrading ICM [4,15]. But our previous study revealed the degradation effectiveness of UV process on the triiodinated compounds [15], which are attributed to iodine element in structure. Besides, UV also exhibits enhanced effect on the formation of chloropicrin, N-nitrosodimethylamine, halobenzoquinones, haloacetic acids and even I-DBPs [15,29–33]. Therefore, it is important to investigate the UV degradation effects of representative ICM-iohexol, on the I-DBPs formation during subsequent oxidation.

The main aim of this research was to investigate the UV photodegradation features of iohexol and the I-THMs formation during subsequent chlorination. The factors of UV fluence, solution pH, Br⁻ concentration as well as the raw water matrix and UV degradation mechanism of the triiodinated compound were also studied. This study could be of some relevance to understand the transformation of nontoxic iodinated organic compounds as ICM to toxic I-DBPs in UV-chlorination processes.

2. Materials and methods

2.1. Chemicals

All chemicals were at least of analytical grade except as noted. Iohexol (CAS No: 66108-95-0) was purchased from Sigma-Aldrich (USA) in more than 99.0% purity and used as received. Iodoform (CHI₃, 99%), sodium hypochlorite (NaOCl) solution (available chlorine 4.00-4.99%), NaOH (≥ 98%), KH₂PO₄ (≥ 99.0%), Na₂CO₃ (≥ 99.0%), NaHCO₃ (≥ 99.0%), KI (\geq 99.0%) were also purchased from Sigma-Aldrich. Five I-THMs standards, including chlorodiiodomethane (CHCll, 90–95%), dichloroiodomethane (CHCl, \geq 95%), bromochloroiodomethane (CHBrClI, \geq 95%), dibromoiodomethane (CHBr,I, 90-95%) and bromodiiodomethane (CHBrI₂, 90–95%) were obtained from CanSyn Chemical Corp. (Canada). Methyl tert-butyl ether (MtBE) and acetonitrile were obtained from J.T. Baker (USA). Formic acid solution (49-51%) was purchased from Fluka (St. Louis, MO, USA) and atrazine was obtained from Dr. Ehrenstorfer (German) in chromatographical purity (>99.0%). Analytical grade reagents including Na₂S₂O₃, H₂O₂, K₂TiO(C₂O₄)₂, H_3PO_4 and H_2SO_4 were purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). All solutions were prepared with ultra-pure water produced from a Milli-Q water purification system (Millipore, USA).

The raw waters of Yangtze River and Huangpu River were collected from Yangshupu drinking water treatment plant (Shanghai, China) and Minhang No. 2 drinking water treatment plant (Shanghai, China), respectively. Then the raw water samples were filtered through 0.45 µm acetate membrane filters (Millipore, USA) immediately and stored in the dark at 4°C until usage. The characteristics of HR and YR were provided in Table 1.

The chemical and physical properties of iohexol and iopamidol were presented and compared in Table 2. It can be seen from Table 2 that, although the iodine content is a bit smaller, iohexol has greater molecular weight and molar extinction coefficient than its analogue. The different chemical and physical properties of iohexol might virtually influence its UV photodegradation features.

2.2. UV irradiation

The photodegradation experiments were conducted using a bench-scale UV quasicollimated beam setup housed with four low pressure Hg UV lamps. The system contained a 9 L stainless steel reactor (i.d. = 20 cm, height = 32 cm) with a 400 mL quartz tube (i.d. = 4.5 cm) fixed in the center. The temperature of the UV system was controlled at 25°C by connecting the system to a low temperature water bath (DKB-1915, Jinghong, Shanghai, China) and circulating the

Table 1	
Characteristics of raw water samples	

Parameters	Yangtze River raw water	Huangpu River raw water
pН	7.29	7.75
Turbidity	9.44	27.5
NH_4 – N (mg/L)	0	0.12
UV ₂₅₄	0.046	0.107
TOC (mg/L C)	2.003	4.261
$Br^{-}(\mu g/L)$	47.22	92.96
TN (mg/L N)	1.732	2.614
NO_3^-N (mg/L)	1.105	0.095

Table 2 Chemical and physical properties of iohexol and iopamidol



water by a diaphragm pump (Model DP0700, Puricom, Taiwan, China). Four UV lamps (TUV 11 W T5 4P-SE, Philips, Netherlands) with quartz sleeves were symmetrically fixed in the center of the reactor. The UV intensity was changed by turning on different numbers of UV lamps and detected using a light intensity meter (UV-C luxometer, Photoelectric Instrument Factory of Beijing Normal University, Beijing, China) at 254 nm. The UV intensity of 1.53, 3.02, 4.45 and 5.96 mW cm⁻², were measured with UV lamps of 1, 2, 3 and 4, respectively. The determination of UV fluence rate values as 6.891×10⁻⁷, 14.159×10⁻⁷, 18.879×10⁻⁷, 25.487×10⁻⁷ einstein s⁻¹ L⁻¹, using atrazine actinometer [34] and path lengths as 7.225, 6.405, 6.616, 6.443 cm using H₂O₂ actinometry [35] were detailed elsewhere in our previous study [15]. The UV fluence (mJ cm⁻²) delivered to the sample was calculated as the UV intensity multiplied by the exposure time [36,37].

2.3. Experimental procedures

The iohexol solutions were beforehand buffered using 10 mM phosphate (for pH 5–8) or 10 mM carbonate (for pH 9) buffer. Solution pH was adjusted with small volume of H_2SO_4 and/or NaOH. Br⁻ was spiked into iohexol solution before exposed to established UV intensity for 5 min. In UV degradation experiment, a 200-mL solution spiked with 10 μ M iohexol and buffered at certain pH with temperature controlled at 25±1°C was subjected to UV irradiation. 1 mL of solution was rapidly transferred into a UPLC vial at different time-points, and then the samples were analyzed by ultra performance liquid chromatography (UPLC) immediately.

The analyses of iohexol photodegradation intermediates experiments were carried out during UV irradiation at a concentration of 25 μ M so as to enhance production for higher analytical resolution. At the scheduled time, the samples were collected and analyzed by UPLC-ESI-MS as soon as possible. In order to reveal the I-THMs formation features in chlorination process after UV irradiation of the samples, a completely removal of iohexol within 10 min UV exposure was applied in this study. After exposed for 10 min at certain UV intensity, the stock chlorine was spiked into solutions to the desired concentration. Then the I-THMs formation experiments were conducted in triplicate under headspace-free conditions in 40-mL glass screw-cap amber vials with PTFE-lined septa in dark at temperature ($25 \pm 1^{\circ}$ C) in a thermostatic biochemical incubator. At the scheduled time of 7 d, the reaction was quenched using NH₄Cl at 1.2 times of oxidants concentration and the samples were then extracted for the I-THMs analysis.

2.4. Analytical methods

The solution pH was measured by a regularly calibrated pH meter (FE20-FiveEasy, Mettler Toledo, Switzerland) using standard buffer solutions (Mettler Toledo; pH = 4.01, 7.00, 9.21). The chlorine concentration was calibrated by the N, N-diethyl-p-phenylenediamine (DPD) colorimetric method [38]. The UV intensity changed by turning on different numbers of UV lamps were measured as 1.53, 3.02, 4.45 and 5.96 mW cm⁻², respectively, by placing a light intensity meter (UV-C luxometer, Photoelectric Instrument Factory of Beijing Normal University, Beijing, China) into a quartz sleeve. The UV absorbance was measured via a 1-cm quartz cell at wavelength of 254 nm using a UV-Vis spectrophotometer (SQ-4802 UNICO, Shanghai).

Iohexol and I⁻ released from photolysis of iohexol were both analyzed using UPLC (Waters, USA) equipped with a XTerra® MS C18 column (4.6 × 250 mm i.d., 5 µm film thickness, Waters, USA) by an UV detector at wavelength of 245 nm and 223 nm [15,39,40], respectively. The mobile phase for iohexol was consisted of 5%/95% (v/v) acetonitrile and Milli-Q water at a flow rate of 0.80 mL min⁻¹. The mobile phase for I⁻ was consisted of 2%/98% (v/v) acetoni-

58

trile and KH_2PO_4 solution (0.09 M) at a flow rate of 0.80 mL min⁻¹. The injection volume was 10 µL. The detection limits of iohexol and I⁻ were 10.0 and 5.0 µg L⁻¹, respectively.

Method for quantifying I-THMs was developed by modifying USEPA Method 551.1 [15,41]. Samples were extracted with MtBE and the extracts were analyzed using a gas chromatograph (GC-2010, Shimadzu, Japan) equipped with an electron capture detector (ECD) and a HP-5 capillary column (30 m \times 0.25 mm i.d., 0.25 μ m film thickness, J&W, USA). The injector and detector temperature were 200 and 290°C, respectively, and the flow rate of the nitrogen carrier gas was 30 mL/min with pressure of 69.8 kPa. The temperature program for the regular DBPs analyses was as follows: hold at 37°C for 10 min, ramp to 50°C at 5°C/ min and hold for 5 min, and finally ramp to 260°C at 15°C/ min and hold for 10 min. The temperature program for the iodinated DBPs analysis method was as follows: hold at 40°C for 10 min and then ramp to 260°C at 15°C/min and hold for 10 min.

Photodegradation intermediates of iohexol during UV irradiation were identified by UPLC-ESI-MS consisted of an Accela U-HPLC system and a TSQ Quantum mass spectrometer (ESI source, Thermo Scientific Inc.,USA). An XTerra® MS C18 column (250 mm \times 2.1 mm, i.d., 5 mm film thickness, Waters, USA) was used for product analysis. Acetonitrile (10%) and 1% formic acid solution (90%) were used as the mobile phase at a flow rate of 0.25 mL min⁻¹. MS chromatograms were obtained both in total ion current (TIC) mode, using full scans (m/z 200–900) for mass spectra acquisition and in the selected ion monitoring (SIM) mode.

3. Result and discussion

3.1. UV degradation kinetics

The UV degradation kinetic model of organic substances has been reported by many researchers and the concentration decrease of the organic compound by UV versus time can be expressed as the following equation [15,26,27,41]:

$$-\frac{dC_{t}}{dt} = \Phi I_{0} \Big[1 - \exp(-2.3L\varepsilon C_{t}) \Big]$$
(1)

Then Eq. (2) can be obtained by integrating Eq. (1):

$$\ln\left(\frac{10^{\varepsilon L C_0} - 1}{10^{\varepsilon L C_t} - 1}\right) = 2.3 \Phi I_0 L \varepsilon t = k_{obs} t \tag{2}$$

The parameters C_0 and C_t indicate the concentrations of the target compound at the UV irradiation time of t = 0 and t = t (M); I_0 is the UV fluence rate value (einstein s⁻¹ L⁻¹); Φ represents the quantum yield of the compound (mol einstein⁻¹); ϵ is the molar extinction coefficient of the compound (M⁻¹ cm⁻¹); *L* is the optical path length (cm). As expressed in Eq. (2), plot of the left term of the equation towards the irradiation time should be a straight line, the slope of which is the pseudo-first order rate constant ($k_{obs} = 2.3 \Phi I_0 L \epsilon$).

Hence, according to the phodegradation results of iohexol, the fitting lines under different UV intensities are shown in Fig. 1.

As displayed in Fig. 1, the linear fits of iohexol photodegradation under different UV intensities were excellent enough to utilize the above kinetic model ($R^2 > 0.98$). More-

0.05 Experimental data 12 0.0 itting line, R²=0.94 0.03 [s.] 10 k obs 0.0 In[(10^{eLc0}-1)/(10^{eLCt}-1)] 8 0.0 0 2 6 $L^{-1}) = 10$ - [UV], = 1.53 mW cm⁻², R² = 0.987 - [UV]₃ = 4.45 mW cm⁻², R² = 0.982 ☆ [UV]₄ = 5.96 mW cm⁻², R² = 0.993 100 150 200 250 300 Time (s)

Fig. 1. Pseudo-first order kinetics plot of iohexol photodegradation under different UV intensity of 1.53, 3.02, 4.45, 5.96 mW cm⁻², respectively. (pH = 7, [phosphate buffer] = 10 mM, [iohexol]₀ = 10 μ M.)

over, faster degradation of iohexol was observed at greater UV intensity. In Fig. 1, this phenomenon corresponds to the increasing slopes of the fitting lines with UV intensity. The results are consistent with other scholars' reports on the photooxidation of other organic materials [15,26–27]. It can also be seen from Fig. 1 that the photodegradation effectiveness of iohexol by UV irradiation was proved, which can be attributed to the photosensitivity of the triiodinated compound. The k_{obs} and iohexol degradation after 5 min UV exposure under different UV intensities are provided in Table 3.

Furthermore, the regression analysis of k_{obs} and UV fluence rate values was also plotted in Fig. 1, the equation of which can be fitted as $k_{obs} = 1592.7 I_0$. Then, according to the ε of iohexol (27620 M⁻¹ cm⁻¹) [3], values of $I_{o'}$ and *L* of the UV reactor, the Φ of iohexol at 254 nm can be calculated as 0.0385 mol einstein⁻¹, which was similar to the former value estimated by Pereira et al. ($\Phi = 0.0403$ mol einstein⁻¹) [3]. The quantum yield of iohexol evaluated in this study was a bit bigger than iopamidol ($\Phi = 0.03318$ mol einstein⁻¹, $\varepsilon = 22700 \text{ M}^{-1} \text{ cm}^{-1}$), which was consistent with their molar extinction coefficient difference. The experimental data also indicated that iohexol showed more removal than iopamidol by UV irradiation.

3.2. Effect of pH and Br on iohexol photodegradation

Study on the photodecomposition of iohexol by UV irradiation was further extended to the effect of solution pH and Br⁻ and the first-order rate constants under different conditions are summarized in Table 3.

As shown in Table 3, the k_{obs} of iohexol photodegradation and degradation percent after 5 min UV exposure were not much affected by the increase of solution pH from 5 to 10. The results indicated that solution pH has no remarkable influence on the UV photodegradation rate of iohexol. The data is directly related to the fact that the undissociated molecule of iohexol in solution would lead to constant

59

Table 3			
First order rate constants of iohe	kol degradation	during UV	' irradiation

Concentration of Br ⁻ (µM)	UV intensity (mW cm ⁻²)	pН	k _{obs} (s ⁻¹)	t _{1/2} (s)	Iohexol degradation after 5 min UV exposure (%)	R ²
0	1.53	7	0.0155	154.4	82.65	0.987
0	3.02	7	0.0212	101.8	97.01	0.991
0	4.45	7	0.0281	78.96	100.00	0.982
0	5.96	7	0.0415	52.29	100.00	0.993
0	3.02	5	0.0215	100.4	99.57	0.972
0	3.02	6	0.0214	100.9	99.21	0.975
0	3.02	7	0.0212	101.8	97.03	0.963
0	3.02	8	0.0213	101.4	98.49	0.956
0	3.02	9	0.0211	102.8	96.87	0.968
0	3.02	10	0.0210	102.8	96.46	0.973
10	3.02	7	0.0213	101.4	98.35	0.993
25	3.02	7	0.0212	101.9	97.03	0.977
50	3.02	7	0.0212	101.9	97.00	0.986
100	3.02	7	0.0214	100.9	98.86	0.988
200	3.02	7	0.0213	101.4	98.41	0.982
0.591 ª	3.02	7	0.0185	116.7	90.56	0.969
1.163 ^b	3.02	7	0.0143	151.0	80.37	0.972

Notes:

^aThe concentration of Br⁻was obtained from the raw water samples of Yangtze River.

^bThe concentration of Br⁻ was obtained from the raw water samples of Huangpu River.

monochromatic absorption coefficient [26]. This is consistent with our previous research on the UV photodegradation of iopamidol [15].

It can also be seen from Table 3 that no significant difference of k_{obs} was observed among different Br⁻ concentrations. As bromide would not transform under UV light from low pressure mercury vapor lamps [43], the increase of Br⁻ concentrations could not change the monochromatic absorption coefficient of the solution and so was the photodegradation rate of iohexol. The Br⁻ separated in waters may not be involved in the UV phototransformation pathways of iohexol. It is also deduced from our experimental results that changing the acidity or basicity and Br⁻ concentrations of waters will not improve the UV degradation efficiency of iohexol.

3.3. UV photodestruction pathways of iohexol

The nuclear magnetic resonance spectra has revealed that the UV-light energy could cause the structural dissolved humic substances change from high-molecular-weight matter to more aliphatic contents with more carboxyl and carbonyl carbon atoms [37]. Previous reports on the degradation of ICM have verified the deiodinated and hydroxylated intermediates during advanced oxidation/reduction processes, simulated solar radiation as well as UV irradiation [15,45,46]. The UV degradation intermediates of iohexol were analyzed by UPLC-ESI-MS and the proposed structures for the products are summarized in Fig. 2. The product of 695 was characterized as substitution of one iodine atom by a hydrogen atom on the aromatic ring of iohexol. The stepwise deiodination of iohexol was demonstrated by the simultaneous degradation of iohexol and release of I⁻ in Fig. 3. Our previous research has proved the stepwise deiodination and hydroxylation of the triiodinated compound by UV irradiation at the iodo sites in benzene ring [15]. Similarly, the ions of 695, 569, 585 and 601 were identified as partially or completely hydroxylated intermediates of three iodo sites in iohexol. Other ions of 819, 693, 709, 583, 599, 489 and 473 were postulated as the ketone products by hydrogen atom abstraction reactions from side chains of the deiodinated intermediates of iohexol [15,45,46]. In general, the UV photodestruction pathways of iohexol were proposed and presented in Fig. 2.

3.4. Effect of UV fluence on I-THMs formation during post-chlorination

Report has proven that the photodegradation of iopamidol by UV irradiation would not be accompanied by the mineralization of total organic carbon (TOC) in solution [15]. Therefore, the photooxidation intermediates of iohexol after UV irradiation might provide organic carbon source for DBPs formation during subsequent chlorination. Meanwhile, I⁻ released from the UV deiodination of iohexol could be active iodine source for the formation of toxic I-DBPs in subsequent oxidation process [10,11]. Thus the I-THMs formation from chlorination of iohexol pretreated by different UV intensities is exhibited in Fig. 4.

60



Fig. 2. Proposed destruction pathways of iohexol during UV irradiation.



Fig. 3. Simultaneous degradation and deiodination of iohexol with UV intensity of 3.02 and 5.96 mW cm⁻², respectively. (pH = 7, [phosphate buffer] = 10 mM, [iohexol]₀ = 10 μ M.)



Fig. 4. Effect of UV fluence on the I-THMs formation during irradiation of iohexol followed by chlorination process. Error bars represent standard deviation of three replications. (UV fluence = 918, 1812, 2670 and 3576 mJ cm⁻², pH = 7, [phosphate buffer] = 10 mM, [iohexol]₀ = 10 μ M, [Cl₂]₀ = 100 μ M, t = 7 d).

It should be pointed out that no I-THMs was measured during the chlorination of iohexol without UV irradiation. However, as shown in Fig. 4, UV irradiation induced the formation of three kinds of I-THMs (including CHCl₂I, CHCll₂ and CHI₃) to the following chlorination process. Furthermore, the increase of UV fluence significantly enhanced the formation of all the detected I-THMs. The UV irradiation of iohexol resulted in the generation of I⁻ in solution and then HOI can be formed after spiking with Cl₂, afterwards I-THMs were formed from the oxidation of the photodegradation products by HOI [13,19], which is shown in Eq. (3) [47]. Although CHCl₂I constituted the majority of I-THMs, the formation of CHI₃ might cause serious taste and odor problems in drinking water as its organoleptic threshold concentration lay as low as 0.03–1 µg L⁻¹ [13,48].

$$HOCl + I^{-} \to HOI + Cl^{-} \quad k_0 = 4.3 \times 10^8 M^{-1} s^{-1}$$
 (3)

$$ISF = \frac{\left[CHCl_{2}I\right] + \left[CHBr_{2}I\right] + \left[CHClBrI\right] + 2 \times \left[CHCll_{2}\right] + 2 \times \left[CHBrl_{2}\right] + 3 \times \left[CHl_{3}\right]}{\left[CHCl_{2}I\right] + \left[CHBr_{2}\right] + \left[CHClBrI\right] + \left[CHClBrI\right] + \left[CHClBrI_{2}\right] + \left[CHBrl_{3}\right]}$$
(4)

Like the application of bromine substitution factor (BSF), iodine substitution factor (ISF) was introduced in this study to provide a direct measure of iodine content of the total halogen incorporated into each group of I-THMs [40, 49]. Therefore, ISF can be calculated by Eq. (4) to compare the iodination degrees of the detected I-THMs with different UV fluence. It can be seen from Fig. 4 that ISF constantly increased with elevating UV fluence, which clearly indicated an enhancive iodine substitution of the total I-THMs formation.

3.5. Effect of pH and Br on I-THMs formation during post-chlorination

As discussed in Section 3.2, pH and Br⁻ posed no distinct effect on the photodegradation rate of iohexol. However, what we care about most is whether these factors would influence I-THMs formation in the following oxidation process. Accordingly, the effect of Br⁻ on I-THMs formation during subsequent chlorination is shown in Fig. 5.

It was found that the effect of pH on I-THMs formation as well as ISF were negligible, which can be attributed to the fact that variations of pH values didn't change the amount of I- formed from the photodegradation of iohexol (data not shown). As shown in Fig. 5, however, the effect of Br- concentration on the I-THMs formation in UV-chlorination of iohexol was distinct. Not only the I-THMs yields but also the species of I-THMs were significantly increased. The introduction of Br- to solution led to the enhanced formation of brominated I-THMs, which increased with the Br- concentrations. Meanwhile, although the sum formation of CHCl₂I and CHCll, was depressed at higher Br- concentrations, the CHI₃ formation was obviously enhanced. As Br⁻ concentrations gradually increased, the total yields of I-THMs considerably elevated and the I-THMs species with more iodine atoms (CHClI₂, CHBrI₂ and CHI₃) increased, which caused increasing ISF values evidently. The results agreed with previous studies concerning Br- effect on I-THMs of different experimental conditions [40,50]. Overall, the increase of ISF primarily resulted from the more powerful oxidation capacity of HOBr and HOI shown as Eqs. (5) and (6) in the presence of Br⁻ [51,52]. Our conclusions also suggested that the naturally occurring Br- in iohexol-containing raw waters, especially for the sea water intrusion areas [11], could highly enhance the formation risk of toxic I-THMs.

$$HOCl + Br^{-} \rightarrow HOBr + Cl^{-} \quad k_1 = 1550 \ M^{-1}s^{-1} \tag{5}$$

$$HOBr + I^{-} \rightarrow HOI + Br^{-} \quad k_2 = 5.0 \times 10^9 M^{-1} s^{-1}$$
 (6)

3.6. Effect of the raw water matrix on iohexol photodegradation and I-THMs formation during post-chlorination

It is known that the presence of natural water components might affect the degradation rate and phototrans-



Fig. 5. Effect of Br⁻ concentration on the I-THMs formation during UV irradiation of iohexol followed by chlorination process. Error bars represent standard deviation of three replications. (UV fluence = 1812 mJ cm⁻², [Br⁻]₀ = 0, 10, 25, 50, 100 and 200 μ M, pH = 7, [phosphate buffer] = 10 mM, [iohexol]₀ = 10 μ M, [Cl₂]₀ = 100 μ M, t = 7 d).

formation of organic compounds by UV irradiation [34]. Accordingly, two raw waters spiked with 10 μ M iohexol were studied for the photodegradation and I-THMs formation during subsequent chlorination. The k_{obs} of iohexol photodegradation and degradation percent after 5 min UV exposure in the two raw water matrix were calculated and are given in Table 3. The results obtained on the effect of raw water matrix on I-THMs formation in UV-chlorination of iohexol are shown in Fig. 6.

It can be inferred from Table 3 that the raw water matrix exhibited a negative effect on iohexol removal rates and the k_{obs} was observed in decreasing order as follows: PW > YR > HR. This is because that UV energy absorbed to iohexol decreased due to the consumption of UV energy by NOM in raw waters [2]. The more severe competition circumstances with NOM for UV energy in HR than YR led to the smaller degradation rate in the latter raw water matrix. Some scholars have also reported that NOM present in natural waters can reduce the effective fluence rate and consequently the rate of photodegradation [2,34]. Our results were consistent with these conclusions.

However, as for the I-THMs formation during subsequent chlorination, the raw water matrix displayed contrarily. As shown in Fig. 6, much more I-THMs were measured in HR and YR, especially brominated I-THMs, which were corresponding to the occurrence of Br⁻ in these two raw waters (Table 1). The distinct higher I-THMs formation observed in real source waters might be due to the fact that NOM are important precursors in the formation of I-DBPs [9,16]. Although the degradation rate and transformation of iohexol in HR were the smallest of all (Table 3), it inhabited comparatively the most I-THMs (Fig. 6) because of its greatest NOM content (Table 1). Therefore, the important roles of NOM on I-THMs formation in UV irradiation and sequential chlorination processes should be concerned. Similar conclusion can also be drawn from



Fig. 6. Effect of the raw water matrix on the I-THMs formation during UV irradiation of iohexol followed by chlorination process. Error bars represent standard deviation of three replications. (UV fluence = 1812 mJ cm⁻², pH = 7, [phosphate buffer] =10 mM, [iohexol]₀ = 10 μ M, [Cl₂]₀ = 100 μ M, t = 7 d, the abbreviates of "PW", "YR" and "HR" represent the water matrix of Pure Water, Yangtze River raw water and Huangpu River raw water, respectively.)

the comparison of ISF increased in the order as PW < YR < HR in Fig. 6. The results revealed that though the NOM presence in actual source waters could weaken the iohexol removal by UV irradiation, it might conversely contribute to the substantial enhancement of toxic I-THMs formation in the following chlorination process.

4. Conclusions

Iohexol can be photodegraded by UV irradiation with pseudo-first order kinetics and the quantum yield was evaluated as 0.0385 mol einstein⁻¹. The degradation rate of iohexol was linearly increased with UV intensity. But pH and Br⁻ both have no remarkable influence on the UV photodegradation rate of iohexol.

Based on the deiodinated and hydroxylated intermediates identified by UPLC-ESI-MS analyses, the destruction pathways of iohexol by UV irradiation were proposed.

The increase of UV fluence and Br⁻ concentrations significantly enhanced the formation of I-THMs and ISF during post-chlorination. However, the effect of pH was negligible.

The raw water matrix exhibited a negative effect on iohexol removal rates and the k_{obs} was observed in decreasing order as follows: PW > YR > HR. Nevertheless, the I-THMs formation and ISF during UV-chlorination of iohexol were significantly promoted due to the presence of NOM and Br⁻ in the raw water samples.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (No. 51308439), the Shaanxi Provincial Natural Science Foundation (Grant No. 2015JM5161), and the Program for Innovative Research Team in Shaanxi Province (Grant No. 2013KCT-13).

References

- C.G. Daughton, T.A. Ternes, Pharmaceuticals and personal care products in the environment: agents of subtle change? Environ. Health. Persp., 107 (1999) 907.
- [2] I. Kim, N. Yamashita, H. Tanaka, Photodegradation of pharmaceuticals and personal care products during UV and UV/ H₂O₂ treatments, Chemosphere, 77 (2009) 518–525.
- [3] V.J. Pereira, H.S. Weinberg, K.G. Linden, P.C. Singer, UV degradation kinetics and modeling of pharmaceutical compounds in laboratory grade and surface water via direct and indirect photolysis at 254 nm, Environ. Sci. Technol., 41 (2007) 1682–1688.
- [4] W. Seitz, J.Q. Jiang, W. Schulz, W.H. Weber, D. Maier, M. Maier, Formation of oxidation by-products of the iodinated X-ray contrast medium iomeprol during ozonation, Chemosphere, 70 (2008) 1238–1246.
- [5] S. Pérez, P. Eichhorn, M.D. Celiz, D.S. Aga, Structural characterization of metabolites of the X-ray contrast agent iopromide in activated sludge using ion trap mass spectrometry, Anal. Chem., 78 (2006) 1866–1874.
- [6] M. Schulz, D. Löffler, M. Wagner, T.A. Ternes, Transformation of the X-ray contrast medium iopromide in soil and biological wastewater treatment, Environ. Sci. Technol., 42 (2008) 7207– 7217.
- [7] M.N. Sugihara, D. Moeller, T. Paul, T.J. Strathmann, TiO₂-photocatalyzed transformation of the recalcitrant X-ray contrast agent diatrizoate, Appl. Catal. B-Environ., 129 (2013) 114–122.
- [8] T.A. Ternes, R. Hirsch, Occurrence and behavior of X-ray contrast media in sewage facilities and the aquatic environment, Environ. Sci. Technol., 34 (2000) 2741–2748.
- [9] S.E. Duirk, C. Lindell, C.C. Cornelison, J. Kormos, T.A. Ternes, M. Attene-Ramos, S.D. Richardson, Formation of toxic iodinated disinfection by-products from compounds used in medical imaging, Environ. Sci. Technol., 45 (2011) 6845–6854.
- [10] C. Jeong, Drinking water disinfection by-products: toxicological impacts and biological mechanisms induced by individual compounds or as complex mixtures, University of Illinois at Urbana-Champaign, 2014.
- [11] S.D. Richardson, F. Fasano, J.J. Ellington, F.G. Crumley, K.M. Buettner, J.J. Evans, A.B. McKague, Occurrence and mammalian cell toxicity of iodinated disinfection byproducts in drinking water, Environ. Sci. Technol., 42 (2008) 8330–8338.
- [12] M.J. Plewa, E.D. Wagner, S.D. Richardson, A.D. Thruston, Y.T. Woo, A.B. McKague, Chemical and biological characterization of newly discovered iodoacid drinking water disinfection byproducts, Environ. Sci. Technol., 38 (2004) 4713–4722.
- [13] Y. Bichsel, U. Von Gunten, Formation of iodo-trihalomethanes during disinfection and oxidation of iodide-containing waters, Environ. Sci. Technol., 34 (2000) 2784–2791.
- [14] S.D. Richardson, C. Postigo, D. Barceló, Emerging organic contaminants and human health, The Handbook of Environmental Chemistry, 2012, pp. 93–137.
 [15] F.X. Tian, B. Xu, Y.L. Lin, C.Y. Hu, T.Y. Zhang, N.Y. Gao, Photan and Phase Physical Science (2019).
- [15] F.X. Tian, B. Xu, Y.L. Lin, C.Y. Hu, T.Y. Zhang, N.Y. Gao, Photodegradation kinetics of iopamidol by UV irradiation and enhanced formation of iodinated disinfection by-products in sequential oxidation processes, Water Res., 58 (2014) 198–208.
- [16] T. Ye, B. Xu, Z. Wang, T.Y. Zhang, C.Y. Hu, L. Lin, N.Y. Gao, Comparison of iodinated trihalomethanes formation during aqueous chlor(am)ination of different iodinated X-ray contrast media compounds in the presence of natural organic matter, Water Res., 66 (2014) 390–398.
- [17] T.Y. Zhang, B. Xu, C.Y. Hu, Y.L. Lin, L. Lin, T. Ye, F.X. Tian, A comparison of iodinated trihalomethane formation from chlorine, chlorine dioxide and potassium permanganate oxidation processes, Water Res., 68 (2015) 394–403.
- [18] T.Y. Zhang, Y.L. Lin, A.Q. Wang, F.X. Tian, B. Xu, S.J. Xia, N.Y. Gao, Formation of iodinated trihalomethanes during UV/ chloramination with iodate as the iodine source, Water Res., 98 (2016) 199–205.
- [19] Y. Bichsel, U. Von Gunten, Oxidation of iodide and hypoiodous acid in the disinfection of natural waters, Environ. Sci. Technol., 33 (1999) 4040–4045.

- [20] T. Ye, B. Xu, Y.L. Lin, C.Y. Hu, L. Lin, T.Y. Zhang, N.Y. Gao, Formation of iodinated disinfection by-products during oxidation of iodide-containing waters with chlorine dioxide, Water Res., 47 (2013) 3006–3014.
- [21] T. Ye, B. Xu, Y.L. Lin, C.Y. Hu, S.J. Xia, L. Lin, N.Y. Gao, Formation of iodinated disinfection by-products during oxidation of iodide-containing water with potassium permanganate, J. Hazard. Mater., 241 (2012) 348–354.
- [22] E.M. Smith, M.J. Plewa, C.L. Lindell, S.D. Richardson, W.A. Mitch, Comparison of byproduct formation in waters treated with chlorine and iodine: relevance to point-of-use treatment, Environ. Sci. Technol., 44 (2010) 8446–8452.
- [23] R.D. Kashinkunti, K.G. Linden, G.A. Shin, D.H. Metz, M.D. Sobsey, M.C. Moran, A.M. Samuelson, Investigating multibarrier inactivation for Cincinnati-UV, by-products, and biostability, J. Am. Water. Works. Ass., 96 (2004) 114–127.
- [24] A.D. Dotson, C.E. Rodriguez, K.G. Linden, UV disinfection implementation status in US water treatment plants, J. Am. Water. Works. Ass., 104 (2012) 77–78.
- [25] Y.S. Shen, Y. Ku, K.C. Lee, The effect of light absorbance on the decomposition of chlorophenols by ultraviolet radiation and UV/H₂O₂ processes, Water Res., 29 (1995) 907–914.
- [26] F.J. Benitez, J.L. Acero, F.J. Real, S. Roman, Oxidation of MCPA and 2, 4-D by UV radiation, ozone, and the combinations UV/ H₂O₂ and O₃/H₂O₂, J. Environ. Sci. Heal., B 39 (2004) 393–409.
 [27] A. Lopez, A. Bozzi, G. Mascolo, J. Kiwi, Kinetic investigation
- [27] A. Lopez, A. Bozzi, G. Mascolo, J. Kiwi, Kinetic investigation on UV and UV/H₂O₂ degradations of pharmaceutical intermediates in aqueous solution, J. Photoch. Photobio. A, 156 (2003) 121–126.
- [28] A. Hassen, M. Mahrouk, H. Ouzari, M. Cherif, A. Boudabous, J.J. Damelincourt, UV disinfection of treated wastewater in a large-scale pilot plant and inactivation of selected bacteria in a laboratory UV device, Bioresource Technol., 74 (2000) 141–150.
- [29] A.D. Shah, A.D. Dotson, K.G. Linden, W.A. Mitch, Impact of UV disinfection combined with chlorination/chloramination on the formation of halonitromethanes and haloacetonitriles in drinking water, Environ. Sci. Technol., 45 (2011) 3657–3664.
- [30] C. Lee, C. Schmidt, J. Yoon, U. Von Gunten, Oxidation of N-nitrosodimethylamine (NDMA) precursors with ozone and chlorine dioxide: kinetics and effect on NDMA formation potential, Environ. Sci. Technol., 41 (2007) 2056–2063.
- [31] Y. Qian, W. Wang, J.M. Boyd, M. Wu, S.E. Hrudey, X.F. Li, UV-induced transformation of four halobenzoquinones in drinking water, Environ. Sci. Technol., 47 (2013) 4426–4433.
- [32] W. Liu, L.M. Cheung, X. Yang, C. Shang, THM, HAA and CNCl formation from UV irradiation and chlor(am)ination of selected organic waters, Water Res., 40 (2006) 2033–2043.
- [33] W. Liu, Z. Zhang, X. Yang, Y. Xu, Y. Liang, Effects of UV irradiation and UV/chlorine co-exposure on natural organic matter in water, Sci. Total Environ., 414 (2012) 576–584.
 [34] S. Canonica, L. Meunier, U. Von Gunten, Phototransformation
- [34] S. Canonica, L. Meunier, U. Von Gunten, Phototransformation of selected pharmaceuticals during UV treatment of drinking water, Water Res., 42 (2008) 121–128.
- [35] F.J. Beltran, G. Ovejero, J.F. Garcia-Araya, J. Rivas, Oxidation of polynuclear aromatic hydrocarbons in water. 2. UV radiation and ozonation in the presence of UV radiation, Ind. Eng. Chem. Res., 34 (1995) 1607–1615.
- [36] F. Yuan, C. Hu, X. Hu, J. Qu, M. Yang, Degradation of selected pharmaceuticals in aqueous solution with UV and UV/H₂O₂, Water Res., 43 (2009) 1766–1774.
- [37] E.J. Rosenfeldt, K.G. Linden, S. Canonica, U. Von Gunten, Comparison of the efficiency of OH radical formation during ozonation and the advanced oxidation processes O₃/H₂O₂ and UV/H₂O₂, Water Res., 40 (2006) 3695–3704.
- [38] APHA, AWWA, WEF, Standard methods for the examination of water and wastewater, 20th ed., American Public Health Association, Washington, DC, 1998.
- [39] P. Sajonz, J. Bookalam, R.A. Miller, Separation of periodate, iodate and iodide on a C-18 stationary phase. Dependence of the retention on the temperature and solvent composition. Monitoring of an oxidative cleavage reaction, Chromatographia, 64 (2006) 635–640.

64

- [40] Z. Wang, Y.L. Lin, B. Xu, S.J. Xia, T.Y. Zhang, N.Y. Gao, Degradation of iohexol by UV/chlorine process and formation of iodinated trihalomethanes during post-chlorination, Chem. Eng. J., 283 (2016) 1090–1096.
- [41] D. Munch, D. Hautman, Method 551.1: Determination of Chlorination Disinfection Byproducts, Chlorinated Solvents, and Halogenated Pesticides/ Herbicides in Drinking Water by Liquid-Liquid Extraction and Gas Chromatography with Electron-Capture Detection, USEPA, Cincinnati, OH, 1995.
- [42] R. Salgado, V.J. Pereira, G. Carvalho, R. Soeiro, V. Gaffney, C. Almeida, A. Oehmen, Photodegradation kinetics and transformation products of ketoprofen, diclofenac and atenolol in pure water and treated wastewater, J. Hazard. Mater., 244 (2013) 516–527.
- [43] N.H. Phillip, E. Gürten, V. Diyamandoğlu, Transformation of bromine species during decomposition of bromate under UV light from low pressure mercury vapor lamps, Ozone Sci. Eng., 28 (2006) 217–228.
- [44] M. Kulovaara, N. Corin, P. Backlund, J. Tervo, Impact of UV₂₅₄-radiation on aquatic humic substances, Chemosphere, 33 (1996) 783–790.
- [45] J. Jeong, J. Jung, W.J. Cooper, W. Song, Degradation mechanisms and kinetic studies for the treatment of X-ray contrast media compounds by advanced oxidation/reduction processes, Water Res., 44 (2010) 4391–4398.
- [46] S. Pérez, P. Eichhorn, V. Ceballos, D. Barceló, Elucidation of phototransformation reactions of the X-ray contrast medium iopromide under simulated solar radiation using UPLC-ESI-QqTOF-MS, J. Mass Spectrum., 44 (2009) 1308–1317.

- [47] J.C. Nagy, K. Kumar, D.W. Margerum, Nonmetal redox kinetics: oxidation of iodide by hypochlorous acid and by nitrogen trichloride measured by the pulsed-accelerated-flow method, Inorg. Chem., 27 (1988) 2773–2780.
- [48] B. Cancho, C. Fabrellas, A. Diaz, F. Ventura, M.T. Galceran, Determination of the odor threshold concentrations of iodinated trihalomethanes in drinking water, J. Agr. Food Chem., 49 (2001) 1881–1884.
- [49] G. Hua, D.A. Reckhow, Evaluation of bromine substitution factors of DBPs during chlorination and chloramination, Water Res., 46 (2012) 4208–4216.
- [50] Z. Wang, B. Xu, Y.L. Lin, C.Y. Hu, F.X. Tian, T.Y. Zhang, N.Y. Gao, A comparison of iodinated trihalomethane formation from iodide and iopamidol in the presence of organic precursors during monochloramination, Chem. Eng. J., 257 (2014) 292–298.
- [51] K. Kumar, D.W. Margerum, Kinetics and mechanism of general-acid-assisted oxidation of bromide by hypochlorite and hypochlorous acid, Inorg. Chem., 26 (1987) 2706–2711.
- [52] R.C. Troy, D.W. Margerum, Non-metal redox kinetics: Hypobromite and hypobromous acid reactions with iodide and with sulfite and the hydrolysis of bromosulfate, Inorg. Chem., 30 (1991) 3538–3543.