

Evaluation of activation parameters of activated carbon from coffee and cocoa seed husk rests: carbon yields and Ni(II) adsorption capacity study

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ABSTRACT

Activated carbon produced by a traditional two-step activation procedure from coffee husks (HAC) and cocoa seed husks (CAC) were used as adsorbents for Ni(II) removal in aqueous solution. Temperature, activation time and added water amount used during the activation process are studied through a model equation designed by a two-level full factorial design. Analysis of variance was used in evaluating AC in multiple response optimization to maximize the yield and adsorption capacity as system responses. The adsorption behaviour of Ni(II) was also evaluated through isotherm models of Langmuir, Freundlich and Langmuir–Freundlich. For both CAC and HAC, the Langmuir–Freundlich model is slightly superior to the Langmuir and Freundlich models. Mostly CAC adsorbs higher amount of Ni(II) than HAC.

Keywords: Adsorption; Ni(II); Multiple response optimization; Activated carbon; Coffee husk; Cocoa seed husk

1. Introduction

Environmental pollution is still growing as urbanization and industrialization have been intensified in the last decades [1]. Environmental contaminations, particularly the contamination of surface and groundwater resources have become a considerable ecological concern, due to the presence of harmful materials (dissolved heavy metals and organic pollutants, waste from factories) in waters [2].

Heavy metals are hazardous contaminants, due to their toxicity and strong propensity to accumulate in the environment

and in the food chain. The treatment of wastewater contaminated with these metals is a primary concern [3,4].

Activated carbons (ACs) are very effective in treating low metal-ion concentration in aqueous solutions [5–9]. Due to their high amount of micropores and mesopores, their great surface area [3], the variety of surface functional groups interacting with the heavy metal ions and even the possibility to increase the adsorption capacity by modifying with other functional groups or using additives [3,10–12], makes these adsorbents interesting candidates in wastewater treatments. The characteristics of the adsorbent significantly affect their adsorption capacity. Generally, the properties of the AC differ depending on the nature of the raw material and the conditions of the activation process [13].

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The agrowaste receives a lot of attention as cheap source for the production of AC and makes this alternative more attractive for small-scale industries [14–17]. Agricultural waste for AC production is mainly coming from shells and stones of the fruits. However, waste resulting from the production of rice, coffee, maize, cocoa, sugarcane and corn gets more and more attention [18]. The preparation of ACs from waste materials using pyrolysis offers economic and environmental advantages [19]. This is the main reason why despite the effectiveness of other adsorbents as clays, ion-exchange resins, zeolite and synthetic composites [20–24] the low-cost ACs are interesting candidates for heavy metals removal.

Nickel is one of the major toxic metals. Several industries, such as electroplating, non-ferrous metals mineral processing, dyeing and steam-electric power plants have contributed to the contamination of different water bodies with nickel [20]. Mineral processing of this element leaves high levels of nickel ions in aquatic environment [4]. Around 40–45 mg/L is determined in the liquor waste from an acid leaching technology. In addition, the Ni(II) salts are known to be carcinogenic. According to the World Health Organization (WHO) general "safe" concentration values should be below 0.2 mg/L [25,26].

In the search of a proper biomass for the production of cost-effective adsorbents for Ni(II) removal, taking into account its price, purity and availability, also the relationship yield–adsorption capacity is of importance. The carbon contents of these products are lower as compared with anthracite or coal. Therefore, the yields of ACs from these precursors are estimated to be lower. Nevertheless, its lower cost gives noteworthy more impact than its lower yield. The high content of volatile matters presents in the biomass is ideal to produce a highly porous structure of ACs [27].

The purpose of statistical designing of an experiment is to gather the maximum amount of relevant information with a minimum cost of time and resources. The factorial design of experiments, combined with statistical methods of data analysis, offers wider and more distinguished information on the system, while conclusions are of better usability [28]. The aim of the present study is to evaluate the effect of activation temperature, time and water amount (used to produce steam) on yield and adsorption capacity of coffee and cocoa seed husks AC in the adsorption of Ni(II) ions by two-level full factorial design analysis of variance (ANOVA) and multiple response optimization. Also, the adsorption characteristics of Ni(II) are studied by three different adsorption isotherm models: Langmuir, Freundlich and Langmuir–Freundlich.

2. Materials and methods

2.1. Chemicals

All reagents used were of analytical grade and purchased from Sigma-Aldrich NV/SA, Belgium. Metal solution was prepared by dissolving NiSO₄·7H₂O in Milli-Q water to obtain a Ni stock solution of 5,000 mg/L. All other solutions were prepared by diluting this stock solution. The pH of the solution was adjusted with 0.1 N NaOH or 0.1 N HCl solution. The concentration of nickel ions was determined using an inductively coupled plasma spectrophotometer employing a Perkin Elmer Optima 3000 DV ICP-AES device with an axial plasma configuration.

2.2. *Raw materials, preparation and production of activated carbon*

Coffee and cocoa seed husks were acquired from the eastern region of Cuba. AC has been prepared from the above materials. Samples are first pyrolyzed in an oxygen-free atmosphere (N₂) in a lab-scale reactor [29]. For each experiment, a known amount of sample is introduced into the reactor. After the reactor is sealed and placed under a stream of nitrogen (2 × 70 mL/min) the reactor is heated with a rate of 10°C/min to 450°C and then held for an isothermal period of 1 h to complete the pyrolysis process. The sample is continuously kept in motion by an Archimedes screw in order to achieve a uniform heat distribution. The reactor is heated up with a special tailored heating mantle, and the temperature is checked using a thermocouple located inside the reactor [29]. During the thermal treatment, the sample is subjected to a thermal cracking and volatilization. The gases that are formed leave the reactor and passed through a condensation unit. The condensed fraction is pyrolytic oil. The formed biochar remains behind in the reactor and the non-condensable gases leave the system. The biochar is activated through physical activation by steam in a second step.

For activation, the biochar is introduced in a horizontal quartz reactor and fixed with two quartz wool plugs. The biochar is heated up under a N_2 atmosphere to a preselected activation temperature with a heating rate of 20°C/min. At a fixed activation temperature, the atmosphere is switched from N_2 to water vapour to complete the activation process for a given activation time [30].

The yield (%) of coffee husks activated carbon (HAC) and cocoa seed husks activated carbon (CAC) is calculated based on the following equation:

$$Y = \frac{m_{\rm AC}}{m_0} \times 100\tag{1}$$

where $m_{AC}(g)$ is the dry weight of the AC and $m_0(g)$ is the dry weight of the biochar.

2.3. Experimental design

The factorial design is widely used in experiments involving several factors where it is necessary to study the joint effect of the factors on a response. The 2^k design provides the smallest number of trials with which *k* factors can be studied in a complete factorial design [31].

The Design Expert Software version 10 was used for the experimental design and data analysis. In this research, a three-factor two-level (high and low or +1 and -1) full factorial design (2³ trials) was used. Two responses (the yield and the adsorption capacity average) were simultaneously optimized by studying the factors: the activation temperature, activation time and added water amount (to produce steam) at the levels shown in Table 1. ANOVA was used to measure the magnitude of effects of factors studied on the system responses.

Table 1

Independent variables and their coded levels for 2^3 factorial design

Independent variables	Code	Range and levels		Coded res	ponses
		-1	+1	Yield (%)	$q_{\rm exp}$
Temperature, °C	А	850	900	R1	R2
Activation time,	В	30	45		
min					
Added water	С	10	15		
amount, mL					

2.4. Optimization of multiple responses using desirability function

Desirability function *D*, which was proposed by Harrington [32], and later optimized by Derringer and Suich [33], is an optimization technique for one or multiple responses and it has been used by many researchers [34–38]. The desirability process contains three stages: (1) predicting responses on the dependent variable by fitting the observed responses using an equation based on the levels of the independent variables, (2) finding the levels of the independent variables that simultaneously produce the most desirable predicted responses on the dependent variables and (3) maximize the overall desirability with respect to the controllable variables [39].

In order to combine the multiple responses in a single function that can be maximized, a desirability function is defined first for each answer $d_i(\hat{y}_i)$. The desirability function $d_i(\hat{y}_i)$ shows the desirability value on a scale of 0 to 1 (lowest desirability to highest desirability) [35]. When the *n* variables are converted in desirability functions, they are combined into an overall desirability function known as global desirability (*D*), using the following equation [40]:

$$D = (d_1^{r_1} \times d_2^{r_2} \times \dots \times d_n^{r_n})^{\frac{1}{\sum_{i}}}$$
(2)

where the r_i is the relative importance among the *n* variables and responses with target *i* = 1, 2, ..., *n*.

Desirability function $d_i(\hat{y}_i)$ takes some of the three ways, depending if the answer should be maximized, minimized or to reach an objective value, within a suitable range of response value settings by $(U_i - L_i)$ where U_i is the upper acceptable value for the response and L_i is the lower value. In addition, if the response has to be maximized, $d_i(\hat{y}_i)$ is described by the following equation [40]:

$$d_{i}(\hat{y}_{i}(x)) = \begin{bmatrix} 0 & \text{if } \hat{y}_{i}(x) < L_{i} \\ \left(\frac{\hat{y}_{i}(x) - L_{i}}{U_{i} - L_{i}}\right)^{s} & \text{if } L_{i} \leq \hat{y}_{i}(x) \leq U_{i} \\ 1 & \text{if } \hat{y}_{i}(x) > U_{i} \end{bmatrix}$$
(3)

where *s* defines the shape of the function and is a power value named "weight", established by the analyst to determine

how significant it is for \hat{y}_i to be close to the maximum. The equation for minimizing $d_i(\hat{y}_i)$ is as follows [40]:

$$d_i(\hat{y}_i(x)) = \begin{bmatrix} 1 & \text{if } \hat{y}_i(x) < L_i \\ \left(\frac{U_i - \hat{y}_i(x)}{U_i - L_i}\right)^t & \text{if } L_i \le \hat{y}_i(x) \le U_i \\ 0 & \text{if } \hat{y}_i(x) > U_i \end{bmatrix}$$
(4)

where *t* is the weight to determine how significant it is for \hat{y}_i to be close to the minimum. When a target value T_i is the best desirable response, the function setting is [40]:

$$d_{i}(\hat{y}_{i}(x)) = \begin{bmatrix} 0 & \text{if } \hat{y}_{i}(x) < L_{i} \\ \left(\frac{\hat{y}_{i}(x) - L_{i}}{T_{i} - L_{i}}\right)^{s} & \text{if } L_{i} \leq \hat{y}_{i}(x) \leq T_{i} \\ 1 & \hat{y}_{i}(x) = T_{i} \\ \left(\frac{\hat{y}_{i}(x) - U_{i}}{T_{i} - U_{i}}\right)^{t} & \text{if } T_{i} \leq \hat{y}_{i}(x) \leq U_{i} \\ 0 & \text{if } \hat{y}_{i}(x) > U_{i} \end{bmatrix}$$
(5)

2.5. Adsorption isotherms

Adsorption isotherm tests are performed using 25 mg of adsorbent dose and 50 mL of Ni(II) solution (10, 20, 30, 40 and 50 mg/L) in an Erlenmeyer flask of 250 mL. The experiments are carried out at $25^{\circ}C \pm 1^{\circ}C$, solution pH of 6 and shake speed 50 rpm, for 24 h, based on a previous study [41]. After each experiment, the solution is filtered and the concentration of Ni(II) is determined.

The amount of Ni(II) adsorbed q_e (mg/g) at equilibrium is calculated using the following equation:

$$q_e = \frac{C_0 - C_e}{m} \times V \tag{6}$$

where C_0 and C_e are the initial and equilibrium concentration of Ni(II), respectively (mg/L), *V* is the volume of the solution (L) and *m* is the weight of the adsorbent used (g). The average of adsorption capacity (q_e) determined is used as system response (Table 1).

Freundlich, Langmuir and Langmuir–Freundlich models are fitted to adsorption isotherm data for equilibrium description. Freundlich isotherm is an empirical model and is based on a multilayer adsorption, with non-uniform distribution of adsorption, heat and affinities over the heterogeneous surface [42,43] and in linear form it is given by:

$$\log q_e = \log K_F + \frac{1}{n} \log C_e \tag{7}$$

where K_F is related with the adsorption capacity and *n* is related to the adsorption intensity [42].

Langmuir isotherm is based on a theoretical model and assumes a monolayer adsorption over an energetically homogeneous adsorbent surface containing a finite number of adsorption sites. It does not take into account interactions between adsorbed molecules [44,45]. It can be represented by the following linear equation:

$$\frac{1}{q_e} = \frac{1}{q_m} + \left(\frac{1}{K_L q_m}\right) \frac{1}{C_e}$$
(8)

where q_m and K_L are constants related to the maximum adsorption capacity (mg/g) and the adsorption energy (L/mg), respectively [46,47].

The Langmuir–Freundlich isotherm equation [48] represents the combination of Langmuir and Freundlich behaviour through:

$$q_{e} = \frac{q_{mLF} (K_{LF} C_{e})^{n_{LF}}}{(K_{LF} C_{e})^{n_{F}} + 1}$$
(9)

where q_{mLF} K_{LF} and n_{LF} are the maximum adsorption capacity (mg/g), the adsorption energy (L/mg) and the adsorption intensity for Langmuir–Freundlich model. To determine the parameters of the model non-linear curve fitting was applied to the data using the Origin 8.1 program.

2.6. Characterization of the adsorbents prepared at optimal conditions

Porous texture analysis has been carried out by N₂ and CO₂ adsorption at -196°C and 0°C, respectively, in an Autosorb iQ apparatus (Quantachrome Instruments). The samples were outgassed overnight at 300°C before N2 adsorption and for 5 h at 300°C before CO₂ adsorption under high vacuum. The specific surface area ($S_{\rm BET}$) was calculated from the N₂ sorption isotherm data using the Brunauer, Emmett and Teller (BET) method. The amount of nitrogen adsorbed at the relative pressure of $p/p_0 = 0.96$ was used to determine the total pore volume (V_T) . The N₂ isotherms in the p/p_0 range from 0.0005 to 0.96 reflect the adsorption that takes place in the mesopores (pores with a width of 2–50 nm) and in the micropores larger than 0.7 nm. The micropore volume ($V_{\rm DR,N2}$) and the average micropore size $(L_{0,N2})$ were estimated by applying the Dubinin– Radushkevich and Stoeckli equations, respectively, to data collected at low pressures ($p/p_0 < 0.015$) [49]. The $V_{\text{DR,N2}}/V_T$ ratio was used to assess the contribution of the micropores to the total pore volume. The quenched-solid density functional theory (QSDFT) analysis [50] was applied to the N₂ adsorption isotherms to determine pore size distribution (PSD).

The CO₂ isotherms at 0°C and low relative pressure $p/p_0 < 0.1$ are assumed to correspond to the adsorption taking place in the narrow micropores in the range of 0.4–0.8 nm (ultramicropores). These isotherm data are used to calculate the ultramicropore volume ($V_{DR,CO2}$) and ultramicropore size ($L_{0,CO2}$) by means of the Dubinin–Radushkevich and Stoeckli equations, respectively [49]. Assuming the presence of slit-shaped ultramicropores, the surface of their walls was determined from the following equation: S_{CO2} (m²/g) = 2,000

 $V_{\rm DR,CO2}/L_{0,CO2}$ [51]. The PSD is also calculated from the CO₂ isotherm data by applying the non-local density functional theory (NLDFT) [52] in order to characterize ultramicropores.

Elemental analysis of carbon (C), nitrogen (N), hydrogen (H), sulphur (S) and oxygen (O) (by difference) is carried out with a Thermo Electron Flash EA1113 element analyzer with BBOT (2,5-bis(5-*tert*-butyl-benzoxazol-2-yl)thiophene with formula $C_{26}H_{26}N_2O_2S$) as standard for calibration. Attenuated total reflectance Fourier transform infrared spectroscopy (ATR-FTIR) measurements are carried out with a Bruker Vertex 70 equipped with a DTGS detector. The dried samples are directly measured in the wave number range from 4,000 to 600 cm⁻¹ at a resolution of 4 cm⁻¹ using a PIKE accessory.

3. Results and discussion

3.1. Designing of experiments and statistical model assessment

Tables 2 and 3 show the experimental results based on 2³ full factorial design for HAC and CAC. The 16 runs represent the possible combination of coded factors in a random way with one replicate. The observed and predicted responses for ACs adsorption capacity and yield for HAC and CAC at the same experimental conditions are given in Tables 2 and 3, respectively.

The effect of the factors (independent variables) and their interactions on both responses can be represented for an empirical relationship expressed by a linear equation. The coded mathematical models obtained from 2^3 full factorial experimental design model and the input variables for both adsorbents are expressed by Eqs. (10)–(13):

$$R1_{HAC} = 42.91 - 7.07A - 3.52B - 2.74C + 0.66AB -1.50AC - 1.29BC + 0.85ABC$$
(10)

Predicted $R^2 = 0.9958$ and adjusted $R^2 = 0.9922$:

$$R2_{HAC} = 50.52 + 0.27A + 0.49B - 0.41C - 0.076AB + 0.29AC + 0.11BC - 0.059ABC$$
(11)

Predicted $R^2 = 0.9562$ and adjusted $R^2 = 0.9179$:

$$R1_{CAC} = 49.99 - 5.05A - 4.51B - 3.90C + 0.14AB + 0.97AC + 1.76BC + 0.28ABC$$
(12)

Predicted $R^2 = 0.9944$ and adjusted $R^2 = 0.9894$:

$$R2_{CAC} = 55.21 + 1.17A + 2.05B + 1.38C - 0.32AB + 0.22AC - 0.57BC - 0.27ABC$$
(13)

Predicted $R^2 = 0.9921$ and adjusted $R^2 = 0.9851$:

The correlation coefficient R^2 is a measure of the quality of the developed models for predicting a response value. The difference in adjusted R^2 (a measure of the amount of variation on the mean explained by the model) and predicted R^2 (a measure of the variation in data explained by the model) should be approximately 0.20 of each other. In the study,

Run no.	Indep	endent va	riables	Responses	Responses								
	(code	d)		R1 (yield, %))		R2 ($q_{e'}$ mg/g)						
	Α	В	С	Observed	Predicted	Residual	Observed	Predicted	Residual				
1	-1	1	-1	49.44	49.19	0.25	51.13	51.34	-0.21				
2	-1	-1	-1	53.98	53.28	0.70	50.55	50.55	0.00				
$3^{(*,1)}$	-1	1	-1	48.95	49.19	-0.24	51.56	51.34	0.22				
4	1	1	-1	37.28	37.66	-0.38	50.95	51.27	-0.32				
5	1	-1	1	34.28	34.91	-0.63	50.39	50.22	0.17				
6	1	1	1	28.39	28.30	0.01	51.11	51.13	-0.02				
7	-1	-1	1	55.93	55.05	0.88	49.00	48.82	0.18				
8	1	-1	-1	42.76	42.50	0.26	50.55	50.55	0.00				
9(*,4)	1	1	-1	38.04	37.66	0.38	51.60	51.27	0.33				
10	-1	1	1	43.18	42.43	0.75	50.27	50.28	-0.01				
$11^{(*,6)}$	1	1	1	28.20	28.30	-0.01	51.16	51.13	0.02				
12	-1	1	1	41.67	42.43	-0.76	50.28	50.28	0.01				
13(*,8)	1	-1	-1	42.24	42.50	-0.26	50.55	50.55	0.00				
14(*,7)	-1	-1	1	54.17	55.05	-0.88	48.64	48.82	-0.18				
15(*,5)	1	-1	1	35.53	34.91	0.63	50.05	50.22	-0.17				
16(*,2)	-1	-1	-1	52.58	53.28	-0.70	50.55	50.55	0.00				

Table 2 Experimental design based on 2^3 full factorial design with one replicate for HAC

(*,#)Replicate of run #, that is, (*,1) replicate of run 1.

Table 3 Experimental design based on 2³ full factorial design with one replicate for CAC

Run no.	Indep	endent va	ariables	Responses	Responses							
	(code	d)		R1 (yield, %))		R2 (q _e , mg/g	R2 ($q_{e'}$ mg/g)				
	Α	В	С	Observed	Predicted	Residual	Observed	Predicted	Residual			
1	1	1	-1	41.89	41.46	0.43	57.71	57.34	0.37			
2(*,1)	1	1	-1	41.03	41.46	-0.43	56.97	57.34	-0.37			
3	1	1	1	40.10	39.66	0.44	58.82	58.88	-0.06			
4	1	-1	-1	53.72	54.27	-0.55	51.81	52.20	-0.39			
5	-1	-1	-1	66.12	66.05	0.07	50.00	50.21	-0.21			
6(*,5)	-1	-1	-1	65.98	66.05	-0.07	50.42	50.21	0.21			
7	-1	-1	1	52.28	53.34	-1.06	53.37	53.13	0.24			
8	-1	1	1	47.60	47.00	0.60	57.54	57.27	0.27			
9(*,7)	-1	-1	1	54.40	53.34	1.06	52.89	53.13	-0.24			
10(*,8)	-1	1	1	46.39	47.00	-0.61	57.01	57.27	-0.26			
$11^{(*,4)}$	1	-1	-1	54.83	54.27	0.56	52.60	52.20	0.40			
12	-1	1	-1	53.48	53.78	-0.30	55.35	55.55	-0.20			
13(*,12)	-1	1	-1	54.07	53.78	0.29	55.75	55.55	0.20			
14(*,3)	1	1	1	39.23	39.66	-0.43	58.95	58.88	0.06			
15	1	-1	1	43.49	44.33	-0.84	57.17	57.09	0.08			
16(*,15)	1	-1	1	45.16	44.33	0.83	57.02	57.09	-0.08			

(*,#)Replicate of run #, that is, (*,1) replicate of run 1.

all the predicted R^2 are in reasonable agreement with adjusted R^2 . The models can be used to navigate the design space.

The normality of the data is checked by plotting a normal probability vs. externally studentized residuals (Fig. 1). As the data points on the plot are approximate to a straight line, the data are normally distributed. Negative signs in Eqs. (10)–(13) show antagonistic effects, while positive signs show synergistic effects. Fig. 2 illustrates the perturbation in the system responses with the change in the levels of the coded factors from a reference point. For the yield % (R1), the increment in the factors studied have a negative effect for both adsorbents, whereas for adsorption



Fig. 1. Normal % probability vs. externally studentized residual: (a) R1 of HAC, (b) R2 of HAC, (c) R1 of CAC and (d) R2 of CAC.

capacity (R2) the increase of the factors A (temperature) and B (time of activation) have a positive effect. The influence of C (water amount) depends on the raw material used for the production of ACs. Its raise has a negative consequence for the adsorption capacity of HAC and the opposite for CAC.

The effect of activation temperature and time on system responses can be explained as follows: as activation temperature and time increase, the development of new pores as a result of volatile matter released and the widening of existing ones enable the increment of adsorption capacity up to the optimal conditions. Nevertheless, this causes a decrease in the yield. The increment of added water amount (as steam) produces the opening of new pores into the carbon structure, but higher flows damage/destroy more severe AC structure/matrix.

The significance of the model effects and their interactions in the system responses were studied using ANOVA. ANOVA is a statistical technique that subdivides the total variation of data into modules related with definite sources of variation, with the objective of testing hypotheses on the parameters of the model [34]. The sum of squares (SS) quantifies its importance in the process and *P* values <0.05 indicate that model terms are significant. As Table 4 shows, the main effects and their interactions have larger impact on R1 than on R2 for both adsorbents. The models, the main effects and their interactions are significant, except for *AB* and *ABC* effects for R1_{CAC} and R2_{HAC}. Among all significant variables the temperature (*A*) has the largest effect on R1 response for HAC and CAC (Figs. 2(a) and (c)). While for R2 response the activation time (*B*) has the major effect for both ACs (Figs. 2(b) and (d)). After neglecting insignificant terms, the resultant models R2_{HAC} and R1_{CAC} can be expressed as follows (based on Table 4):

$$R2_{HAC} = 50.52 + 0.27A + 0.49B -0.41C + 0.29AC + 0.11BC$$
(14)

$$R1_{CAC} = 49.99 - 5.05A - 4.51B - 3.90C + 0.97AC + 1.76BC$$
(15)

For $R1_{HAC}$ Eq. (10) and for $R2_{CAC}$ Eq. (13) still hold.



Fig. 2. Perturbation on the system responses by the change of the levels of the factors A, B and C: (a) and (b) HAC, (c) and (d) CAC.

3.2. Desirability function for multiple response optimization

Optimization of system responses (yield and adsorption capacity) is carried out by a multiple response optimization or a global desirability function (D). The optimal operation conditions as activation temperature, time and water amount are settled in the range studied to maximize the yield (R1) and the adsorption capacity (R2). The weight for both responses is fixed to 1. The Design Expert Software version 10 allows varying the importance of system responses between 5 and 1 (maximum and minimum values). However, main effects have larger impact on R1 than R2 and when R2 importance rise from 1 to 5 at fixed importance of R1 equal to 5, the adsorption capacity (R2) increases but the yield (R1) decreases. The R1 values decrease more sharply when R2 importance is fixed among 3-5 (Supplementary material). For the present study, the importance of system responses was therefore fixed to 5 and 2 for R1 and R2, respectively.

For HAC, the best desirability value is 0.821 at predicted system responses of R1: 53.11 and R2: 50.58. The operation conditions are temperature 850°C, activation time 30.5 min and water amount 10 mL (Figs. 3(a) and (b)). While for CAC, the best desirability value is 0.603 with predicted system responses of R1: 58.72 and R2: 53.40 at temperature 850°C,

activation time 39.0 min and water amount 10 mL (Figs. 3(c) and (d)). Fig. 3 shows the graphical desirability for both adsorbents.

3.3. Isotherm studies

Adsorption equilibrium tests are crucial to assess adsorption behaviour and develop mathematic equations for design purposes. In Fig. 4, the adsorption isotherms are presented for the ACs prepared at the optimal experimental conditions. The adsorption models of Langmuir, Freundlich and Langmuir–Freundlich (Table 5) are adjusted to equilibrium data. The Langmuir–Freundlich model showed a better fit for both ACs, followed by the Langmuir model. Maximum adsorption capacities q_{mLF} are clearly higher for CAC than for HAC which is in agreement with multiple response optimization results: predicted system response R2 (adsorption capacity) is greater for CAC than for HAC.

3.4. Characterization of the adsorbent materials

Fig. 5(a) depicts the nitrogen adsorption–desorption isotherms at –196°C for the samples prepared at the optimal experimental conditions. For CAC, the isotherm shape

Table 4	
Analysis of variance (ANOVA) results

Adsorbent	Source	R1 (%)				R2 (mg/g))		
		Sum of	Standard	F value	P value	Sum of	Standard	F value	P value
		squares	error			squares	error		
HAC	Model	1,200.12	0.20	273.65	< 0.0001	9.34	0.058	24.97	< 0.0001
	Α	800.61	0.20	1,277.85	< 0.0001	1.20	0.058	22.43	0.0015
	В	198.25	0.20	316.42	< 0.0001	3.78	0.058	70.78	< 0.0001
	С	120.56	0.20	192.43	< 0.0001	2.67	0.058	50.01	0.0001
	AB	6.92	0.20	11.04	0.0105	0.093	0.058	1.74	0.2236
	AC	35.76	0.20	57.08	< 0.0001	1.36	0.058	25.39	0.0010
	ВС	26.57	0.20	42.41	0.0002	0.18	0.058	3.38	0.1033
	ABC	11.46	0.20	18.29	0.0027	0.055	0.058	1.03	0.3392
CAC	Model	1,044.52	0.22	201.58	< 0.0001	128.66	0.090	142.84	< 0.0001
	Α	408.75	0.22	552.19	< 0.0001	21.90	0.090	170.21	< 0.0001
	В	325.71	0.22	440.01	< 0.0001	67.32	0.090	523.19	< 0.0001
	С	243.91	0.22	329.50	< 0.0001	30.69	0.090	238.52	< 0.0001
	AB	0.33	0.22	0.44	0.5245	1.64	0.090	12.73	0.0073
	AC	15.00	0.22	20.26	0.0020	0.80	0.090	6.23	0.0372
	ВС	49.60	0.22	67.00	< 0.0001	5.15	0.090	40.05	0.0002
	ABC	1.24	0.22	1.67	0.2321	1.16	0.090	8.98	0.0172

Note: The bold values in the table refer to the proposed models, where the main effects and their interactions are insignificant, being thus for both ACs the models AB and ABC.



Fig. 3. Desirability 3D response surface plot: (a) The effect of temperature and water amount for HAC desirability at fixed activation time of 30.5 min, (b) The effect of temperature and activation time for HAC desirability at fixed water amount of 10 mL, (c) The effect of temperature and water amount for CAC desirability at fixed activation time of 39.0 min and (d) The effect of temperature and activation time for CAC desirability at fixed water amount of 10 mL.

shows a sharply increase at low relative pressure of p/p_0 and reaches a plateau in a broad range of p/p_0 . This shape could be classified as a Type I isotherm, characteristic of microporous materials, having mainly narrow micropores [53]. For HAC, the isotherm shape increases continually until the end of relative pressure, indicating the presence of mesopores. Furthermore, the isotherms of both ACs display a hysteresis loop characteristic for Type IV isotherm which confirms the contribution of mesopores to their porous structure. The lack of the lower closure point in the hysteresis loop could suggest these are ink bottle-shaped mesopores [54]. However, the micropores are predominant in the studied ACs as shown in Table 6. The V_{DR}/V_T ratio is much higher for CAC than that for HAC (0.67 vs. 0.81). Both ACs show comparable surface area (S_{BET}) but HAC has higher total pore volume than CAC (Table 6). The PSD determined by QSDFT method reveals a more intense maximum at a pore width of 0.57 nm for CAC and HAC (Fig. 5(b)). Others peaks centre at 0.79, 1.01, 1.54 nm for CAC and 0.85, 1.54 nm for HAC are also found. Based on the N₂ adsorption isotherms,



Fig. 4. Adsorption isotherms for CAC and HAC ($C_0 = 10-50$ mg/L, m = 25 mg, V = 50 mL, pH = 6, $T = 25^{\circ}$ C ± 1°C, shake speed 50 rpm and t = 24 h).

the total pore volume (V_{τ}) is determined at a relative pressure of $p/p_0 = 0.96$ and corresponds to the sum of micropore volume (V_{DR}) and mesopore volume (V_{mes}) . In the context of adsorptive properties of ACs, micropores are crucial because most contaminants are adsorbed in micropores. Only for removing molecules of the size higher than the width of



Fig. 5. (a) Nitrogen adsorption–desorption isotherms at -196° C and (b) QSDFT pore size distribution.

Table 5 Adsorption isotherms parameters (25 mg of AC, 50 mL of Ni(II): $C_0 = 10-50$ mg/L, $T = 25^{\circ}C \pm 1^{\circ}C$)

Conditions		Langmuir model			Freundlich model			Langmuir–Freundlich model			
(uncoded variables)		$q_m (\mathrm{mg/g})$	K_L (L/mg)	R^2	K_{F}	п	R^2	$q_{mLF} (mg/g)$	$K_{\rm LF}({\rm L/mg})$	$n_{\rm LF}$	R^2
<i>A</i> : 850°C, <i>B</i> : 30 min, <i>C</i> : 10 mL	HAC	102.72	0.2918	0.9692	27.17	2.20	0.9221	81.33	0.4632	1.63	0.9870
<i>A</i> : 850°C, <i>B</i> : 39 min, <i>C</i> : 10 mL	CAC	104.99	0.4547	0.9912	34.95	2.33	0.9623	110.15	0.4061	0.93	0.9916

Table 6

Textural parameters for HAC and CAC

Sample	$S_{\rm BET}$ (m ² /g)	V_T (cm ³ /g)	$V_{\rm DR,N2}$ (cm ³ /g)	$V_{\rm DR,N2}/V_T$	$L_{0,N2}^{*}(nm)$	$V_{\rm DR,CO2}$ (cm ³ /g)	$S_{0,CO2}(m^2/g)$	L _{0,CO2} * (nm)
HAC	438	0.250	0.168	0.67	0.84	0.166	976	0.34
CAC	428	0.204	0.165	0.81	0.81	0.140	778	0.36

 $L_0 = 10.8/(E_0 - 11.4)$, where E_0 is the characteristic energy [45].

micropores, ACs with developed mesoporosity are recommended. Macropores (with a width larger than 50 nm) play only a role of transporting arteries.

L is the average pore width, which is calculated by applying the Stoeckli equation: $L_0 = 10.8/(E_0 - 11.4)$ [45] to N₂ adsorption isotherm data ($L_{0,N2}$) and CO₂ adsorption isotherm data ($L_{0,CO2}$), where E_0 is the characteristic energy.

The prepared ACs are analyzed by sorption of CO_2 to characterize ultramicropores. Fig. 6 shows the CO_2 adsorption isotherms (a) and the PSD determined by NLDFT method (b) in the range of ultramicropores. The ACs show a bimodal distribution of ultramicropores size with a maximum centred at 0.35 nm, the second maximum centre is located at 0.52 and 0.54 nm for CAC and HAC, respectively. The surface area and volume achieved for the narrow micropores are greater for HAC than for CAC (Table 6).

The elemental compositions of adsorbent materials at the optimal experimental conditions are shown in Table 7. The carbon and oxygen content for HAC are higher than for CAC, but the nitrogen content for CAC is a little bit higher than for HAC. However, the ATR-FTIR spectra referring to oxygen containing functional groups are more intense for CAC than for HAC (Fig. 6). This partly explains the somewhat better adsorption performance of CAC towards Ni(II) than HAC, also a higher amount of nitrogen functionalities can benefit CAC more than HAC in adsorption performance towards Ni(II). The amount of sulphur is below detection limit for both samples. The literature survey shows the

Table 7 Elemental composition

Adsorbent material	Elemen	Elemental composition (wt%)							
	С	Н	Ν	S	O*				
HAC	59.30	1.34	0.75	0.00	13.69				
CAC	48.82	1.21	0.84	0.00	11.61				

*By difference: O% = (100 - C - H - N - ash) %.

Table 8

Comparison of ACs surface area and adsorption capacity with other adsorbents

Adsorbent	q _m (mg/g) from Langmuir model	S _{BET} (m²/g)	Solution pH	Dosage (g/L)	Concentration range (mg/L)	Reference
Lotus stalks derived activated carbon	31.00	1,220	_	1.0	20-40	[57]
Calcium-sepiolite	4.81	-	-	1.0	5-100	[20]
Tetraethylenepentamine Rosa Canina-L	128.21	-	6	5.0	5-500	[9]
fruits activated carbon						
Mesoporous graphitic carbon nitride	19.39	164	-	1.0	10-100	[58]
$(mpg-C_3N_{4/1.5})$						
Activated carbon from rubber tires	9.34	465	_	0.1	0.1-40	[59]
Activated carbon from cherry kernels	77.71	657	6	2.0	5-500	[60]
Modified magnetic chitosan chelating resin	40.15	55	5	1.5	50-400	[61]
Synthetic PVA/NaX nanofibres	342.80	212	-	0.5	50-1,000	[22]
Activated carbon from coffee husks, HAC	102.72	438	6	0.5	10-50	Present
						study
Activated carbon from cocoa husks, CAC	104.99	428	6	0.5	10-50	Present
						study

size distribution.

maximum adsorption capacities from Langmuir model and the surface area of several adsorbents (Table 8). It is noticed that there is no linear relation between higher surface area and maximum adsorption capacities. Even when the surface





Fig. 7. ATR-FTIR spectra for -HAC and -CAC.

area of both HAC and CAC adsorbents are lower than some of the studies reported [57,59,60] achieved higher or comparable adsorption capacities toward Ni ions.

The ATR-FTIR spectra (Fig. 7) for HAC shows the bands located at 3,639 cm⁻¹ correspond with isolated hydroxyl group –OH stretch [55]; 1,373 cm⁻¹ is characteristic of phenol or tertiary alcohol –OH stretch [55]; the several peaks between 700 and 610 cm⁻¹ can be attributed to aromatic C–H and –OH out of plane bend [9]. CAC has peaks at 3,245 cm⁻¹ (hydroxyl group, H–bonded –OH stretch) [55], 1,511 cm⁻¹ (aromatic C=C–C vibration stretch) [55], 1,385 cm⁻¹ (methyl –CH₃ group) [55], 1,038 cm⁻¹ (primary alcohol C–O stretch or C–C skeletal vibration) [55,56] and several peaks among 900–620 cm⁻¹ (aromatic C–H and –OH out of plane bend) [9].

4. Conclusions

The effect of temperature, activation time and water amount (used to produce steam) on yield and Ni(II) ions adsorption capacity of coffee and cocoa seed husks activated carbons (HAC and CAC) was successfully studied through a two-level full factorial design, ANOVA and multiple response optimization. For the yield (R1), the increment in the factors studied has a negative effect for both adsorbents, whereas for adsorption capacity (R2) the increase of the factors A (temperature) and *B* (time of activation) has a positive effect. The influence of C (water amount) depends on the raw material used for the production of ACs. The main effects and their interaction have larger impact on R1 than R2. The models, the main effects and their interactions are significant, except the interaction of AB and ABC effects. Multiple response optimization technique for maximizing the yield and adsorption capacity gave higher values of R1 and R2 for CAC than the ones achieved for HAC at the optimal temperature, activation time and water amount for each adsorbent.

Isotherm data for the ACs prepared at the optimal experimental conditions is described best by the Langmuir– Freundlich model, followed by the Langmuir model. The maximum adsorption capacities achieved were higher for CAC than for HAC. CAC seems to be more suitable for the removal of Ni(II) than HAC.

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186

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Supplementary material

Table S1 Predicted system responses for HAC at different important values

Importance of syste	em responses	Desirability	Predicted values of system responses				
R1 (yield, %)	R2 ($q_{e'}$ mg/g)		R1 (yield, %)	R2 ($q_{e'}$ mg/g)	A (°C)	B (min)	<i>C</i> (mL)
5	1	0.855	53.28	50.55	850	30.0	10
5	2	0.821	53.11	50.58	850	30.5	10
5	3	0.813	50.02	51.18	850	42.0	10
5	4	0.823	49.20	51.34	850	45.0	10
5	5	0.832	49.20	51.34	850	45.0	10

Table S2 Predicted system responses for CAC at different important values

Importance of syst	nportance of system responses		Predicted values of system responses						
R1 (yield, %)	R2 ($q_{e'}$ mg/g)		R1 (yield, %)	R2 ($q_{e'}$ mg/g)	A (°C)	B (min)	C (mL)		
5	1	0.677	59.98	51.98	850	35.0	10		
5	2	0.603	58.72	53.40	850	39.0	10		
5	3	0.579	56.30	54.45	850	41.9	10		
5	4	0.575	54.40	55.28	850	44.2	10		
5	5	0.579	53.78	55.55	850	45.0	10		