A review on the significance of *Azolla* for water and wastewater treatment

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ABSTRACT

Industrialization, urbanization, and population growth release many pollutants into water resources. Conventional mechanical wastewater treatment plants require a lot of electricity, are not sufficient to remove all pollutants, and generate a large amount of sludge. Therefore, it is crucial to develop economically accepted and environmentally friendly methods for wastewater treatment. The main advantages of bioremediation over conventional treatment methods include low cost, high efficiency, and minimizing the generation of chemical or biological sludge. Aquatic macrophytes can improve water quality by bioaccumulation/biosorption of toxic substances, excess nutrients and metals. Therefore, phytoremediation as a natural system presents itself as an environmentally friendly treatment technology for water remediation. This paper presents recent research on *Azolla* fern as a phytoremediation agent. *Azolla* can remove many pollutants from wastewater, including fluoride, ammonium, bisphenol A, dyes, chemical oxygen demand, nitrogen, phosphorus and toxic metals. *Azolla* is a floating water fern and its biomass produced during phytoremediation has various uses, such as animal feed, human food, production of hydrogen fuel, biogas production, medicine, water treatment, and biological fertilizer in rice fields. Therefore, phytoremediation with *Azolla* is recommended as an advanced treatment after conventional wastewater treatment.

Keywords: Azolla; Bioaccumulation; Fern; Macrophytes; Wastewater treatment

1. Introduction

In 2020, about two billion people still lack access to safely managed water, while about 3.6 billion people worldwide have inadequate sanitation. In addition, about 44% of household wastewater was discharged without safe treatment in 2020. By 2030, current progress must be quadrupled to achieve universal access to safely managed water, sanitation, and basic hygiene services [1]. Therefore, it is crucial to develop economically feasible and environmentally friendly methods for wastewater treatment [2]. Aquatic plants growing in or near the water, known as macrophytes, can be used as a suitable polisher when nutrient concentrations are low [3]. Toxic metals enter the aquatic environment from metal refining, mining, smelting, battery manufacturing, tanneries, petroleum refining, paint manufacturing, pesticides, pigment manufacturing, printing and photographic industries, etc. [4]. In addition, other pollutants such as excess nutrients (especially nitrogen and phosphorus) from livestock watering, fertilizer application, animal feed and animal metabolic residues are also released into aquatic ecosystems [5]. Eutrophication occurs when high concentrations of nutrients, especially nitrogen and phosphorus, enter surface waters [6]. Nutrient removal by expensive methods such as electrocoagulation [7], electrochemical precipitation [8], anaerobic/anoxic/aerobic (A²/O), and biological aerated filter (BAF) has been widely studied [9]. These mechanical methods are expensive and use electricity to accelerate treatment processes that occur naturally [10]. Reducing energy consumption and operating costs of wastewater treatment is a major concern even for small communities in developed countries [11]. Therefore, it is important to develop

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innovative methods to increase treatment efficiency while reducing overall costs. Phytoremediation, as a natural water reclamation system, is an environmentally friendly approach that uses plants as purification agents [12]. Known advantages of using macrophytes for wastewater treatment include simplicity, good biosorption potential, low operating costs, availability, and ease of propagation [4,13]. In addition, biosorption methods offer a good cost-benefit ratio for wastewater treatment and often provide better results than chemical methods [14]. Aquatic macrophytes such as *Azolla* improve water quality due to their great ability to remove organic and inorganic pollutants [15]. Fig. 1 illustrates some of the many possible applications of *Azolla* [16,17].

Based on morphological and reproductive aspects, there are seven obvious species of *Azolla*, which are divided into two subgenera: *Euazolla* and *Rhizosperma*. *Euazolla* includes *A. caroliniana*, *A. filiculoides*, *A. mexicana*, *Azolla microphylla* and *A. rubra*, while *Rhizosperma* includes *A. pinnata* and *A. nilotica* [18]. *A. filiculoides* Lam. which grows in Europe, North America, South Africa, Australia, and New Zealand [19] has advantages such as a high growth rate and the ability to absorb pollutants [20]. Therefore, it is a suitable candidate for phytoremediation [12].

Researchers investigated the ability of *Azolla filiculoides* to remove fluoride [21], ammonium [15], bisphenol A [22], dye [23], chemical oxygen demand (COD) [24], nitrogen, phosphorus, zinc ions and heavy metals (e.g., Pb, Cd, Ni and Cu) from wastewater [6,25–27]. The results of the previous researches as well as the review articles show that the use of *Azolla* can be a suitable and cost-effective option for phytoremediation of polluted water [28]. In this review, most of the capabilities of *Azolla* in water reclamation are presented. In addition, the bioaccumulation and biosorption capacity of *Azolla*, the mechanisms of sorption processes, and the potential of using the biomass produced are discussed.

2. Geographical distribution and characteristics of Azolla

Azolla is a native to the tropics, subtropics, and warm temperate regions of Africa, Asia, and America [29]. This is a small-leaf floating fern (Fig. 2).

Azolla is commonly found in stagnant or slow-moving water in ponds, lakes, marshes, swamps, and streams. Azolla leaves turn red under stress, such as nutrient deficiency, salinity or high temperatures [31]. Under favorable conditions, Azolla rapidly produces biomass (90.0-97.2 kg·dryweight ha⁻¹·d⁻¹) with high protein yields (208–176 g·kg⁻¹) without the need for nitrogen fertilization [32]. This unique freshwater fern grows rapidly in N-free medium (relative growth rate greater than 0.5 d⁻¹) [33]. Due to its symbiotic relationship, Azolla/Anabaena fixes atmospheric nitrogen and releases it as NH4+ in leaf cavities, which is taken up by ferns to produce biomass [34,35]. Colonies of Anabaena/ cyanobacteria are found in the apical meristem within the megasporangium indusium and in an ovoid cavity on the adaxial side at the base of each dorsal leaf-lobe of the Azolla leaf [19]. The studies carried out on A. filiculoides have shown that the growth rate in real wastewater and in artificial media with low nitrate concentration or even without nitrogen does not show a significant difference (0.122-0.126 d⁻¹). A high concentration of ammonium ions in a mineral medium has an inhibitory effect on the *Azolla/Anabaena* association [33]. It is noteworthy that the high ammonium concentration in artificial media (34–40 mg·N·L⁻¹) has a negative effect on the growth and nitrogen fixation of *A. filiculoides*, but this effect was not observed in real wastewater [36]. Although *Azolla* has several advantages, it is sometimes considered an invasive plant in wetlands because dense *Azolla* mats shade submerged macrophytes [29,37]. Fig. 3 shows the floating *Azolla* mat on the water surface.

The availability of water is a crucial factor for the growth of *Azolla*. Optimal growth conditions are light intensity (15–18 K Lux), water temperature (18°C–28°C), relative humidity (55%–83%), and the presence of macro- and micro-nutrients. Wind and turbulent water can fragment and kill *Azolla* [29]. *Azolla* is not able to grow sustainably in a saline



Fig. 1. Azolla applications.



Fig. 2. Azolla fern in the water [30].



Fig. 3. Floating Azolla mat on the surface of water [30].

medium. Salinity has a strong negative influence on photosynthesis of *Azolla*. Moreover, *Azolla/Anabaena* symbiosis in nitrogen fixation is also affected by the negative effect of salinity [38]. Six *Azolla* species (*A. filiculoides, A. mexicana, A. microphylla, A. pinnata, A. rubra,* and *A. caroliniana*) were studied in saline growth medium by adding 0.32% NaCl under polyhouse conditions. The results showed that *A. microphylla* and *Azolla rubra* were more tolerant to salinity. It was also found that *A. microphylla* performed better than the other species in terms of biomass productivity [39].

3. Mechanisms of Azolla for the degradation of pollutants

Bioremediation is a controlled process that uses microorganisms to degrade hazardous organic pollutants to harmless compounds such as CO₂, CH₄, and H₂O [40]. Phytoremediation is a bioremediation process that uses plants and their associated microorganisms to remove pollutants from water or soil [41]. The four main mechanisms of phytoremediation are shown in Table 1 [42]. Azolla can uptake metals, organic and inorganic pollutants from the aquatic ecosystem, either in the form of living biomass (bioaccumulation) or inactive/dead biomass (biosorption) [13,43]. Plants generally use several extracellular and intracellular mechanisms to remove heavy metals. The external mechanism is the exudation of phytosiderophores into the rhizosphere. The exudation mechanism can chelate and dissolve metals bound to the soil. Two ways to enhance metal accumulation in plant tissue are acidification of the rhizosphere and ion exchange sites on cell walls to bind metal ions. Metals are bound by the cell wall, transport systems, and intracellular mechanisms for uptake across the plasma membrane [43].

The root surface (rhizosphere) of *Azolla* hosts a microbiome that can support successful phytoremediation of metals. The identification of rhizobacteria in the root zone of *A. filiculoides* L. treated with metals (Pb, Cd, Cr(VI), Ni, Au, Ag) reveals the presence of known metal-tolerant genera (Mucilaginibacter, Pseudomonas, Mycobacterium, Corynebacterium, Stenotrophomonas, Clostridium, Micrococcus, Achromobacter, Geobacter, Flavobacterium,

Table 1 Phytoremediation processes and mechanisms of contaminant removal [42]

Process	Mechanism	Contaminant
Rhizofiltration	Rhizosphere	Organics/
	accumulation	inorganics
Phytostabilization	Complexation	Inorganics
Phytoextraction	Hyper-accumulation	Inorganics
Phytovolatilization	Volatilization	Organics/
	by leaves	inorganics
Phytotransformation	Degradation in plant	Organics

Arthrobacter, and Delftia) [44]. Rhizofiltration is an effective sub-process of phytoremediation that takes place in the roots of plants and can reduce concentration of heavy metals in polluted waters [45]. In phytoremediation, the interaction between plants and their associated microorganisms causes the release of various metabolites (e.g., 1-aminocyclopropane-1-carboxylic acid deaminase, indole-3-acetic acid, siderophores, organic acids, etc.). These compounds with acidifying, chelating and/or reducing effects facilitate phytoremediation through the processes of translocation, transformation, chelation, immobilization, solubilization, precipitation, volatilization and complexation of heavy metals [46]. A comparison of the rhizofiltration capacity of Azolla, Pistia (water lettuce) and Eichhornia (water hyacinth) in reducing the initial concentration of As (5,000 µg·L⁻¹), Cu (25 mg·L⁻¹), Pb (66 mg·L⁻¹), Ni (58.16 mg·L⁻¹) and fluoride (50 mg·L-1) was performed. The results showed that Eichhornia had the greatest effect in reducing Pb and As, so that the final concentrations of these two elements were 0.189 ± 0.007 and 12.683 ± 1.002, respectively. Azolla had the best performance in absorbing nickel, while Pistia had a greater effect in reducing the initial concentration of copper, and the final concentration of these two elements in water was 1.479 ± 0.032 and 0.265 ± 0.036 , respectively. These observations were associated with an increase in the content of organic acids (citric acid and malic acid) as root exudates, so that Nickel treated with Azolla significantly increased the content of these 2 acids by 1.7 times [47].

4. Removal of pollutants from aqueous media by Azolla ferns

As there are many kinds of pollutants in the environment and many technologies to deal with them. In this situation, we summarized a comprehensive and instructive review based on the previous published literatures of phytoremediation of various *Azolla* species in aqueous solutions for removing organic and inorganic contaminants (Table 2).

According to Table 2, *Azolla* species, living or non-living biomass, single/multiple pollutant solution, solution type, chemical properties of the pollutant, initial pollutant concentration, and contact time mainly affect the removal efficiency. It was reported that light intensity, water temperature, pH, macro- and micronutrient availability, and water turbulence were the most important factors affecting the growth of *Azolla*. Thus, *Azolla* shows higher removal efficiency in

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Species of Azolla	Type of medium	Pollutants	Removal rate	Condition	References
Azolla filiculoides	Aqueous solution	Pb^{2+} , Cd^{2+} , Nl^{2+} , and Zn^{2+}	Inactivated <i>Azolla</i> with methanol: Pb ²⁺ : 36%, Cd ²⁺ : 33%, Ni ²⁺ : 34% Zn ²⁺ : 24% Inactivated <i>Azolla</i> with ethanol: Pb ²⁺ : 41%, Cd ²⁺ : 36%, Ni ²⁺ : 38% and Zn ²⁺ : 31%.	100 mL of the heavy metals solution (25 mg/L for each metal ions) were prepared individually. The solutions were incubated with 2.0 g (dry $Azolla$)/L and pH 5.5 ± 0.2 at 22°C ± 2°C with orbital shaking (150 rpm). After 1 h exposure, metals-laden biomass was separated by centrifugation	[14]
Azolla filiculoides	Olive mill wastewater (OMWW)	COD and polyphenols	Traditional extraction systems (TS) of OMWW: COD: 52%, polyphenols: 53% Continuous extraction systems (CS) of OMWW: COD: 95%, polyphenols: 65%	- 5:1 OMWW to $Azolla$ -fresh-weight ratio - COD (mg/L) in TS 92,000 ± 2,200 and in CS 44,400 ± 800 - Total polyphenols (mg/L) in TS 7,360 ± 290 and in CS 4,367 ± 130 - Fresh $Azolla$ biomass (200 g), the OMWW volumes 1 L, contact time 30 min	[24]
Azolla pinnata	Aqueous solution	Acid Blue 25	36.5 ± 0.5% at the fifth cycle; 41.2 ± 3.6% at the first cycle.	 - A 500 mg/L AB 25 stock solution - C_i ranged from 40 to 500 mg/L and temperature (35°C, 45°C, 55°C and 65°C) - Both adsorbents were treated with 100 mg/L AB 25 of adsorbent dosage 2.0 g/L - pH 2.0, and short duration of contact time at 180 min 	[48]
Azolla pinnata	Water body (Lake)	Heavy metals	Cu (97.7%), Pb (79.7%), Cr (74%), Cd (85.7%) and Zn (97.1%)	 10 g of healthy plants, exposed to 4 mg/L concentration of 5 different heavy metals 10 d experimentation period During experimental period pH 7.0 ± 0.2, water temperature 25°C ± 2.0°C Five different concentrations of DWW including 0% 25% 	[49]
Azolla pin- nata R.Br.	Dairy wastewater (DWW)	pH, electrical conductivity, total dissolved solids, total Kjeldahl's nitrogen, and total phosphorus	9.41%, 61.42%, 71.56%, 73.25%, and 65.37%	50%, 75%, and 100% were used - The average temperature, humidity, and light intensity of the experimental room were 20°C, 55%–70%, and 2,000 lux, respectively - Experimental meriod: 12 h for 14 d	[50]
Azolla pin- nata R.Br.	Integrated industrial effluent (IIE)	TDS, BOD, COD, TKN, Ca, Mg, Na, K, MPN, SPC, P, Cd, Cu, Cr, Fe, Pb, and Zn	25.1, 54.1, 68.8, 71.8, 74.1, 81.9, 45.0, 30.0, 20.9, 41.9, 60.0, 54.1, 60.0, 57.3, 53.9, 58.1, 56.1, 72.4, and 60.0	 Effluent samples were collected from each concentration of treatment pot at the 0, 7, 14, 21, and 28th days during the phytoremediation experiments 	[51]
Azolla filiculoides	Wastewater	Ni and Cu	Maximum uptake capacities for Ni 0.77 and Cu 0.54 mmol/g (dry <i>Azolla</i>)	 - 0.5 to 1 mm size particles of Azolla - Experiments were carried out at room temperature (25°C±1°C) 	[20]
Azolla filiculoides	Aqueous solution	Acid Green 3 (AG 3) dye	99.1% in optimum conditions	- Optimum conditions: $PH = 3$, contact time = 90 min, adsorbent dosage = 4 g/L and dye concentration = 10 mg/L	[52]

 Table 2

 Summarizes the uptake of various pollutants by Azolla reported in the literature

Table 2 (Continued)

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Table 2					
Species of Azolla	Type of medium	Pollutants	Removal rate	Condition	References
Azolla/ Anabaena	Freshwater fish breed- ing areas	Ammonia	Highest removal rate was 6.394/h at light intensity of 140 µmol/m²-s and at a temperature of 28°C; the lowest was 0.947/h at 20 µmol/m²-s and 15°C	- Under artificial light of 20, 70, and 140 μmol/m²·s - Water temperatures for the growing <i>Azolla</i> (15°C, 22°C, and 28°C)	[53]
Azolla filiculoides	Industrial wastewater	Cr(V1)	83.3% at pH of 2	30 mL of different chromium concentration solutions, 1 h of contact time, with 0.1 g of <i>Azolla filiculoides</i> biomass	[54]
Azolla filiculoides	Aqueous solution	Bisphenol A	More than 90% when BPA concentra- tion was 5 ppm and biomass was 0.9 g	 Containers with the capacity of 200 mL Desired concentration of BPA including 5, 10, 25, and 50 ppm Azolla mass was used 0.3, 0.6, 0.9 and 1.2 g Contact time 2, 8, 12, 16, and 20 d 	[22]
Azolla microphylla with papaya stem	Pond waste- water	Nitrate, ammonia, and phosphate	57%, 62%, and 71%, Respectively	100 g of <i>Azolla</i> with 100 g of papaya stem, contact time 24 d	[55]
Azolla filiculoides	Petroleum refinery wastewater	N and P	In separate medium: 36% and 44%, respectively In mixed solution of these two com- pounds: 33% and 40.5%, respectively	- Volume of wastewater: 1 L - Mass of <i>Azolla</i> : 24 g - Contact time 28 d	[34]
Azolla microphylla	Municipal effluents	Total organic C, total Kjeldahl's N, and total P	42%, 39% and 17%, respectively	0.5 g of <i>Azolla</i> , 200 mL of Espinase and Watanabe medium, contact time 7 d	[56]
Azolla pinnata	Aqueous solution	Cu, Pb, Cr, Cd and Zn	44%, 3.94%, 2.85%, 6% and 34%, respec- tively	Azolla were exposed to 4 mg/L concentration of different heavy metals, contact time 10 d, 5 L trays	[57]
Azolla pinnata	Industrial effluents	$\mathrm{Hg}^{2^{+}}$ and $\mathrm{Cd}^{2^{+}}$	70%-94%	Contact time 13 d, volume of aquariums: 40 L, 58.5 g Azolla added to each aquarium	[58]
Azolla pinnata	Reservoir water	Hg	80%-94%	Contact time 6 d, volume of glass aquariums: 40 L, volume of <i>Azolla</i> : 210 g/aquarium	[59]
Azolla pinnata	Aqueous medium	Cr(III)	70%-88%	Contact time 13 d, aquariums capacity: 40 L, 58.5 g <i>Azolla</i> added to each aquarium	[60]
Azolla filiculoides	Aqueous solutions	Acid Blue (AB) 15 dye	98%	200 mg/L initial AB 15 concentration, contact time 10–180 min, 100 mL solution, optimum condition (pH 3, contact time 90 min, adsorbent dose 10 g/L and AB 15 con- centration 10 ppm	[61]
Azolla filiculoides	Industrial wastewater	Reactive Red 198	97.3%	 Initial dye concentration was 10, 25, 50, 100, and 200 ppm Absorbent dosage: 0.2–1.4 g Contact time: 10, 20, 30, 45, 60, 90, 120, 180 and 240 min bH: 3, 5, 7, 9 and 11 	[62]

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Table 2					
Species of Azolla	Type of medium	Pollutants	Removal rate	Condition	References
Azolla filiculoides	Wastewater	Au (gold)	98.2% from a gold Plating factory containing 5 mg·gold/L in solution, 100% of gold from diluted wastewater, 98% of gold from undiluted wastewater containing 41 mg·Au/L	- Stock gold solution 1,000 mg/L - Providing contact times of 60 or 20 min, respectively.	[63]
Azolla filiculoides	Aqueous solution	Phenol	up to 95% at 30°C, more than 97% when phenol concentration was 5 ppm and biomass was 0.9 g	Capacity of each container 200 mL, concentrations of phenol solution were 5, 10, 25, and 50 ppm, <i>Azolla</i> was used in certain weights including 0.3, 0.6, 0.9 and 1.2 g, measured on days 4, 8, 10, 12, 16, and 14	[64]
Azolla filiculoides	Aqueous solution	Fluoride	98%	Optimum pH was 5, contact time up to 75 min, fluoride concentration of 10 mg/L	[65]
Azolla filiculoides	Aqueous solution	COD, TP, and TN	77.5%, 66.8%, and 78.1%	 Synthetic wastewater prepared in the typical range of municipal wastewater (TP 10.8–84.6 mg/L, TN 20–99 mg/L and COD 66.26–415 mg/L) 2 g fresh Azolla floated in 500 CC prepared wastewater Contact time 21 d 	[66]
Azolla	Wastewater	COD, Cd, Pb, Ni, total phosphorous, and total nitrogen	29.68%–98.82%, 92.84%, 97.12%, 76.82%, 46.45%, and 25.57%	 Number of days are between 0 to 30 d Contact time and biomass of <i>Azolla</i> for heavy metal analysis are 15 d and 0.8 g Initial concentration for COD, Cd, Pb, and Ni are 48–4,326, 5, 10, and 25 mg/L 	[67]
Azolla pinnata	Industrial effluents	Hg and Cd	70%94%	 Duration of investigation was 13 d and plant biomass was 58.5 g Initial doses of heavy metals were 0.5, 1, 3 mg/L 	[68]
Azolla filiculoides	Aqueous solutions	Bisphenol A (BPA)	%06-%09	 Biomass level: 0.3, 0.6, 0.9, and 1.2 g Initial concentration of BPA: 5, 10, 25 and 50 ppm Samples were collected every 2 d 	[69]
Azolla pinnata	Palm oil mill effluent	BOD, COD, TS, TSS, oil and grease, NH ₃ -H, and phosphate	63.67%, 80.02%, 85.89%, 80.58%, 81.50%, 98.44%, 92.92% and 0.90 %	- Quantity of <i>A. pinnata</i> : 20–30 g - Concentration of palm oil mill effluent: 10%–60% - Retention time: 1–5 d	[20]

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a solution with one pollutant than in a solution with multiple pollutants. However, in a real water body, there is a mixture of pollutants.

4.1. Removal of inorganic pollutants

The extent of bioaccumulation of pollutants in macrophytes varies greatly. The phytoremediation capacity of duckweed (Lemna minor, Lemna trisulca, Salvinia natans) and water fern (A. filiculoides) was compared. The results show that duckweed species uptake more K, Mg and Na than water fern. A. filiculoides showed lower capacity to bioaccumulate metals than S. natans and L. trisulca. The content of Cd, Cr, Cu, Fe, Ni, and Pb in A. filiculoides was lower than in L. trisulca but higher than in L. minor [71]. Furthermore, the bioaccumulation capacity in plants is metal dependent. In another study, the phytoremediation capacity of L. minor L., A. filiculoides Lam. and Pistia stratiotes L. in contact with Cr, Cd, Pb and Zn was investigated. It was reported that L. minor and P. stratiotes had the highest absorption of Pb, while A. filiculoides absorbed more Cr. In general, the removal efficiency of the aquatic plants studied was in this order: P. stratiotes > L. minor > A. filiculoides [72]. The bio-concentration factor (plant tissue concentration/water concentration) and translocation ability (root concentration/ shoot concentration) of Cd, Cu, Pb, and Ni in three groups of hydrophytes (free-floating, attached, and submerged plants) were studied. It was found that the free-floating plants had better bio-concentration factor and translocation ability than the attached hydrophytes. This is due to the greater surface area to volume ratio of the free-floating plants [73]. The results of a study conducted to investigate the ability of Azolla pinnata to reduce iron in mine wastewater showed that the average value of iron translocation was 1.02. This value shows that the iron concentration is higher in the roots than in the canopy. Iron is taken up by the epidermal cells of the root and enters the xylem vessels of the roots through the apoplast or symplast pathway [74]. Plants synthesized phytochelatins (PCs) upon exposure to heavy metal ions. Phytochelatins are small cysteine-rich peptides that can bind metals via SH groups and play a key role in homeostasis and detoxification of metals and metalloids in plants. Phytochelatins complexes with different toxic metals have different structures, and they are also stable and less toxic than free metal ions [75]. The study of the phytoremediation potential of different Azolla species upon bioaccumulation of Cd, Cu, Zn and Ni showed that the level of metallothionein and phytochelatin synthase encoding genes expression depends on the type of metal, the duration of exposure and the metal concentration in the medium. Since the bioaccumulation of heavy metals and gene expression differ among Azolla species, it can be concluded that an appropriate species should be used for the elimination of the specific metal [76]. The presence of metals in water affects the growth of Azolla. It appears that bioaccumulation with living Azolla biomass cannot continue at high concentrations of toxic metals. Therefore, in highly toxic media, biosorption with inactive/dead biomass seems to be a more sustainable option [77].

Bioremediation of a solution contaminated with hexavalent uranium (U(VI)) followed by real liquid organic radioactive waste was performed with macrophytes (Limnobium laevigatum and Azolla sp.). In the solution containing a range of U(VI) (0.25-36 mmol·L⁻¹), Azolla showed a higher sorption capacity (0.474 mmol·g⁻¹) compared to L. laevigatum (0.026 mmol·g⁻¹). When Azolla was contact with real liquid organic radioactive waste, the sorption capacity decreased to 0.010 mmol·g⁻¹. Therefore, it was recommended to use both macrophytes simultaneously [78]. Although Azolla is able to hyperaccumulate heavy metals, some heavy metals can suppress the growth of Azolla. The extent of the metal effect on growth reduction depends on the type of metal and the Azolla species. Azolla caroliniana was cultivated in a nutrient solution containing metals (Hg²⁺, Cr^{3+} and CrO_4^{2-}). The results show that the presence of these ions inhibits the growth of A. caroliniana by 20%-31% [79]. Different pollutants have different potential to be taken up by roots. For example, the concentration of various heavy metals (Cr, Pb, Hg, Cu, Zn, Cd, Ag and Ti) in the water body and in A. filiculoides was investigated. It was reported that the highest and lowest amounts of heavy metals in Azolla samples were Zn and Cd, respectively. This is because Zn is a vital element for the growth of A. filiculoides [80].

According to previous studies, *Azolla* has been shown to remove heavy metals such as Pb, Cd, Ni, Zn, Cr, Cu, and Hg from aqueous solutions at rates of 4%–41%, 6%–94%, 34%–38%, 24%–34%, 3%–83%, 44%–88%, and 70%–94%, respectively. In the case of nutrients such as nitrogen (total forms) and phosphate, the removal efficiency reached 36%–62% and 17%–71%, respectively. In addition, *Azolla* could remove gold and fluoride up to 98% from aqueous media.

4.2. Removal of organic pollutants

Macrophytes have different potential for phytoremediation of organic pollutants. The removal of organic pollutants was investigated using water hyacinth. The water hyacinth studied can uptake mesotrione and fomesafen (96.7%–98.2%), naphthalene (100%), formaldehyde (93%), sulphadiazine (83.5%) and oxytetracycline hydrochloride, chlortetracycline hydrochloride and tetracycline hydrochloride (>80) [81]. In other study, under optimal conditions (6.3 d contact time, 11.9 g biomass and 14.7 mg/L tetracycline concentration), Azolla removes more tetracycline (100%) compared to chloramphenicol (70%). The mean bio-concentration factors (BCFs) of tetracycline and chloramphenicol were 4.9% and 37.8%, respectively, in Azolla [82]. A. filiculoides was used as a carbon source for the production of activated carbon (ACAF) and for the biosorption of ampicillin from water. The results confirm the promising abilities of Azolla. Under optimal conditions (ACAF dose = 0.8 g/L, pH = 7, ampicillin concentration = 100 mg/L, contact time = 60 min and temperature = 45°C), a removal efficiency of 96.84% (114.3 mg/g) was obtained. The experiments showed that the presence of different concentrations of NaNO₃ competitors did not significantly affect the reduction of Azolla adsorption capacity [83]. In a laboratory-scale phytoremediation study, Azolla was shown to degrade antibiotics in three sequential steps. The first step is the uptake of antibiotics and the release of reactive oxygen species (ROS). In the second step, the effects of ROS are neutralized and minimized by conjugation with enzymes such as glutathione transferase or metabolites such as glutathione. The last step ends with the storage of the assimilated compounds in the vacuoles, the apoplast and the cell wall [84].

Moreover, the chemical properties of water play a crucial role in achieving higher efficiency in the phytoremediation process. For example, one researcher pointed out that the phytoremediation efficiency of water hyacinth significantly depends on the N content [85]. A study on treatability of grey water with water hyacinth showed significant reduction in COD (51.61% ± 13.56%), ammonium-nitrogen (62.15% ± 9.08%), phosphate-phosphorus (58.13% ± 15.23%), turbidity (90.68% ± 7.01%) and total dissolved solids $(9.21\% \pm 2.65\%)$ [86]. The presence of metals can limit or enhance the growth of macrophytes. Some organic substances can also have a toxic effect on Azolla. The study of the toxic effects of bisphenol A (BPA) on A. filicu*loides* showed that BPA ($\geq 20 \text{ mg} \cdot \text{L}^{-1}$) has a negative effect on the growth indices of the Azolla (number of branches, leaf area, growth rate of the plant with severe leaf damage, membrane peroxidation, and electrolyte loss) [87]. As an aquatic fern, A. filiculoides has a wide range of removal efficiency for COD (52%-95%) and polyphenols (53%-65%). It has been reported that A. filiculoides can remove 60%-97% of phenols from an aqueous solution. The reason for the difference in removal efficiency can be explained by the different initial concentrations of phenol or initial Azolla mass [24]. In another case, the researchers showed that the removal of phenol was significantly improved by increasing the temperature, decreasing the initial concentration of the pollutant, and increasing the initial Azolla mass [64]. In a successive sorption-regeneration study, Azolla was shown to remove 36%-99.1% of the Reactive Red dye 198. Removal efficiency depends on the number of regeneration cycles, pH, contact time, plant mass, and initial dye concentration. There is also evidence that 42% of the total organic carbon was removed by Azolla [62]. Evaluation of the phytoremediation ability of A. filiculoides Lam. in nitrogen-free Hoagland culture medium containing 25, 50, and 75 mg·L⁻¹ naphthalene showed that 94% of Azolla was absorbed, accumulated, and/or biodegraded after 10 d [88]. Studies have shown that Azolla has the ability to remediate water contaminated with phenanthrene at concentrations less than 10 mg/L [89].

Comparison of the use of A. filiculoides and Lemna minuta in the treatment of wastewater at concentrations of 5 mg·L⁻¹ for each metal (Fe(III), Cr(VI), Al(III)) and 1 μ g·L⁻¹ for the pharmaceuticals (diclofenac and levofloxacin) shows the superiority of Azolla in removal efficiency and tolerance to pollutants. The removal efficiencies of levofloxacin and diclofenac were 60% and 10%, respectively. Although Cr, Fe and diclofenac had a minor toxic effect on Lemna, the presence of Al had a growth-promoting effect on both plants [90]. A. pinnata and L. minor were used for phytoremediation of palm oil mill effluent (POME) at different dilutions (2.5%, 5%, 10% and 15%). The effluent from this industry is non-toxic in nature and contains a large amount of nutrients (198 mg·L⁻¹ COD, 4.3 mg·L⁻¹ nitrate, pH 9.53, 4 mg·L⁻¹ phosphate, 2.98 mg·L⁻¹ ammonia). Compared to Lemna, Azolla has shown higher efficiency in removing all pollutants within 10 d. Azolla and Lemna have shown significant COD removal efficiencies of 78% and 66%, respectively at a dilution of 15%. The maximum growth of Azolla (296%) was observed at a dilution of 2.5%, but the maximum growth of *Lemna* (535%) was observed at a dilution of 5%. Both plants studied have the ability to absorb nutrients from wastewater and convert them into carbohydrates and proteins for their growth. In Azolla, the amount of carbohydrates in 5% dilution was equivalent to 2 mg·L⁻¹. The carbohydrate content of Azolla increased by 43.71% from the second day to the 10th day. The protein content of Azolla at 2.5% dilution was 26%. Although diluted POME is a suitable medium for the growth of these two plants, increasing the proportion had a decreasing effect on the concentration of carbohydrates and proteins. The results of this study show the ability of Azolla to treat POME with non-toxic and nutritious substances and to produce qualified biomass for animal feedstock [91].

5. Possibilities of using Azolla biomass

In recent decades, increasing concern about the environmental impact of fossil fuels has focused attention on the use of biofuels. On the other hand, the raw materials for biofuels are many plants such as corn and soybeans, which can be used directly for human consumption or indirectly as animal feed [92]. Ethical and economic criticism of biofuel production mostly focuses on the demand for land, water and other resources, which can threaten human food security, increase food prices [93] and even lead to more greenhouse gas emissions [94]. These challenges have drawn attention to the further development of biofuel production technologies and the use of microalgae as biomass [95]. After phytoremediation, the harvested Azolla biomass can be converted into bioenergy in the form of bio-diesel, bio-hydrogen, bio-methane, bio-ethanol and bio-oil as a renewable energy source [96]. A theoretical study to find natural solutions to reduce the risks of fossil fuel consumption has shown that *Azolla* has the ability to sequester 21,266 kg/ha of CO₂ (C) per year from the air and convert it into new biomass [97]. The oil content of Azolla is not as high as that of microalgae, but the problems in separating Azolla biomass from water are much less than with microalgae. The oil extracted from Azolla for biodiesel production is acceptable compared to other feedstock's, and the cost of propagating Azolla biomass for biodiesel production is lower compared to growing the plants [28]. In one study, Azolla was used to treat tannery effluent. Subsequently, Azolla oil was extracted and used to produce biodiesel. The results showed that transesterification of Azolla oil to biodiesel in an optimized process (catalyst loading 2%, molar ratio 1:9, temperature between 60°C and 65°C) is cost effective and reliable [98]. The biomass of A. filiculoides obtained during phytoremediation can be used as a raw material for biofuel production. The ability of A. filiculoides $(11.7 \times 10^3 \text{ L/}$ ha-y) to produce ethanol has been shown to be equivalent to that of corn stover (13.3 \times 10³ L/ha-y). A. filiculoides, with an efficient C/N ratio of 15 (41% total carbohydrates and 20% proteins), is capable of producing biohydrogen at the rate of 2.2 mol H₂/mol reducing sugars [99]. Biofuel is one of the best options for securing future energy supplies from an agriculture-based economy. Because of the high oil content of Azolla, its biomass can be used to produce biogas and biodiesel [100]. Azolla can proliferate in wastewater and produce large amounts of biomass [99]. A. pinnata has been studied as a potential source for biodiesel production. The calorific value (38.2 MJ/Kg) and dynamic viscosity (4.3 cP) of the synthesized biodiesel were similar to those of normal diesel (44 MJ/Kg and 3.06 cP) [101]. Studies on bio-methane production with Azolla biomass grown in water with three different metal concentrations (1, 4 and 8 mg/L) showed that these metals can have different effects. Iron and manganese had no effect on bio-methane production from anaerobic fermentation, but copper, cobalt, lead, and zinc were toxic. At low concentrations, cadmium and nickel have a positive effect on biogas production and methane content, but at high concentrations (8 mg/L) methane production decreases [102]. Review of research on the effect of heavy metals (chromium, cadmium and nickel) in the processes of anaerobic digestion and biogas production from biomass has led to the conclusion that the intensity of inhibition of heavy metals depends on the type of metal and its concentration in dissolved and ionic form in the digester. in addition, the toxic effects of heavy metals on biogas production have been attributed to the disruption of enzyme function and structure by the binding of metal ions to thiol and other groups on protein molecules or by the replacement of naturally occurring metals in the prosthetic groups [103].

Azolla biomass can also be used as animal feed. The dry matter of Azolla species contains proteins (21%-26%), fatty acids (41%-66%), and a wide range of active compounds such as phenols, caffeoylquinic acid derivatives, tannins, and carotenes. The predominant fatty acids in Azolla are palmitic, linolenic, and lignoceric acids [87, 104], so it is logical that Azolla is considered a sustainable source of animal feed in livestock, poultry, and fish diets. On the other hand, Azolla can accumulate heavy metals such as Cr, Pb, Zn, Hg, Cu, Cd, Ag, and Ti [80]. It has been demonstrated that feeding Azolla to Cyprinus carpio growing on water fertilized with sheep manure containing heavy metals (28.5 mg/kg Pb and 139.3 mg/kg Zn) causes histopathological changes and tissue damage in fish [105]. In addition, Azolla is a medicinal plant for sore throat and cough in New Zealand and Tanzania. Azolla may be effective in curing gram-positive bacterial infections [106].

6. Conclusion and recommendations

This review shows that the study of *Azolla* as an aquatic fern in phytoremediation processes has attracted the attention of scientists. Various research articles show that *Azolla* can be used to remove various pollutants in wastewater, such as excess nutrients, heavy metals, dyes, COD, and fluoride. It is a low-cost natural sorbent available in many regions of the world. In addition, after phytoremediation, *Azolla* can be converted into a nutrient-rich organic fertilizer, making phytoremediation with *Azolla* an environmentally friendly approach to removing pollutants.

Consequently, phytoremediation with *Azolla* is recommended as an advanced treatment after conventional wastewater treatment due to its high efficiency, low cost and environmental friendliness in water remediation. However, further studies are needed to assess the use of Azolla as a phytoremediation agent and its impact on real water bodies. In addition, the potential of Azolla for phytoremediation needs to be improved to successfully remove new pollutants. The rapid growth rate and high protein and fatty acid content make Azolla used as animal feed. In phytoremediation with Azolla, the proliferating excess biomass, which may contain a high concentration of pollutants, must be properly disposed of. However, when Azolla is grown in polluted water or wastewater, its use as feed is of concern due to bioaccumulation of pollutants. Therefore, bioenergy production using Azolla biomass grown in wastewater containing toxic metals or organic compounds harmful to human and animal health appears to be a safer option. Most of the research has been conducted on a laboratory scale, which may not correspond to the expected results under real conditions. It is therefore recommended to conduct studies under real conditions.

Declarations

Ethical approval: Not applicable Consent to participate: Not applicable Consent to publish: Not applicable

Authors contributions

Samaneh Taghilou: Data curation, Methodology, Writing – original draft;

Mazyar Peyda: Project administration, investigation, Supervision, Writing – review & editing;

Mohammadreza Mehrasbi: investigation, Writing – review & editing.

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Conflict of interest

The authors of this article declare that they have no conflict of interests.

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References

- [1] WHO, World Health Statistics 2022: Monitoring Health for the SDGs, Sustainable Development Goals, World Health Organization, Geneva, 2022.
- [2] S. Wacławek, H.V. Lutze, K. Grübel, V.V. Padil, M. Černík, D.D. Dionysiou, Chemistry of persulfates in water and wastewater treatment: a review, J. Chem. Eng., 330 (2017) 44–62.

- [3] Y. Tang, S.F. Harpenslager, M.M.L. Van Kempen, E.J.H. Verbaarschot, L.M.J.M. Loeffen, J.G.M. Roelofs, A.J.P. Smolders, L.P.M. Lamers, Aquatic macrophytes can be used for wastewater polishing but not for purification in constructed wetlands, Biogeosciences, 14 (2017) 755–766.
- [4] W.W. Ngah, M. Hanafiah, Removal of heavy metal ions from wastewater by chemically modified plant wastes as adsorbents: a review, Bioresour. Technol., 99 (2008) 3935–3948.
- [5] R. Mithra, S. Sivaramakrishnan, P. Santhanam, S. Dinesh Kumar, R. Nandakumar, Investigation on nutrients and heavy metal removal efficacy of seaweeds, *Caulerpa taxifolia* and *Kappaphycus alvarezii* for wastewater remediation, J. Algal Biomass Util., 3 (2012) 21–27.
- [6] C. Forni, J. Chen, L. Tancioni, M.G. Caiola, Evaluation of the fern *Azolla* for growth, nitrogen and phosphorus removal from wastewater, Water Res., 35 (2001) 1592–1598.
- [7] Y. Tian, W. He, X. Zhu, W. Yang, N. Ren, B.E. Logan, Energy efficient electrocoagulation using an air-breathing cathode to remove nutrients from wastewater, J. Chem. Eng., 292 (2016) 308–314.
- [8] H. Huang, P. Zhang, Z. Zhang, J. Liu, J. Xiao, F. Gao, Simultaneous removal of ammonia nitrogen and recovery of phosphate from swine wastewater by struvite electrochemical precipitation and recycling technology, J. Cleaner Prod., 127 (2016) 302–310.
- [9] Y. Chen, B. Li, L. Ye, Y. Peng, The combined effects of COD/N ratio and nitrate recycling ratio on nitrogen and phosphorus removal in anaerobic/anoxic/aerobic (A²/O)-biological aerated filter (BAF) systems, Biochem. Eng. J., 93 (2015) 235–242.
- [10] S. Rezania, S.M. Taib, M.F.M. Din, F.A. Dahalan, H. Kamyab, Comprehensive review on phytotechnology: heavy metals removal by diverse aquatic plants species from wastewater, J. Hazard. Mater., 318 (2016) 587–599.
- [11] R. Hamza, M.F. Hamoda, M. Elassar, Energy and reliability analysis of wastewater treatment plants in small communities in Ontario, Water Sci. Technol., 85 (2022) 1824–1839.
- [12] F. Vafaei, A.R. Khataee, A. Movafeghi, S.Y.S. Lisar, M. Zarei, Bioremoval of an azo dye by *Azolla filiculoides*: study of growth, photosynthetic pigments and antioxidant enzymes status, Int. Biodeterior. Biodegrad., 75 (2012) 194–200.
- [13] S. Rezania, M. Ponraj, A. Talaiekhozani, S.E. Mohamad, M.F.M. Din, S.M. Taib, F. Sabbagh, F.M. Sairan, Perspectives of phytoremediation using water hyacinth for removal of heavy metals, organic and inorganic pollutants in wastewater, J. Environ. Manage., 163 (2015) 125–133.
- [14] M. Khosravi, R. Rakhshaee, M.T. Ganji, Pre-treatment processes of *Azolla filiculoides* to remove Pb(II), Cd(II), Ni(II) and Zn(II) from aqueous solution in the batch and fixed-bed reactors, J. Hazard. Mater., 127 (2005) 228–237.
- [15] N. Muradov, M. Taha, A.F. Miranda, K. Kadali, A. Gujar, S. Rochfort, T. Stevenson, A.S. Ball, A. Mouradov, Dual application of duckweed and *Azolla* plants for wastewater treatment and renewable fuels and petrochemicals production, Biotechnol. Biofuels, 7 (2014) 30, doi: 10.1186/1754-6834-7-30.
- [16] G. Abraham, D.W. Dhar, Induction of salt tolerance in *Azolla microphylla* Kaulf through modulation of antioxidant enzymes and ion transport. Protoplasma, 245 (2010) 105–111.
- [17] W. Cheng, Y. Okamoto, M. Takei, K. Tawaraya, H. Yasuda, Combined use of *Azolla* and loach suppressed weed *Monochoria vaginalis* and increased rice yield without agrochemicals, Org. Agric., 5 (2015) 1–10.
- [18] U. Kumar, A.K. Nayak, P. Panneerselvam, A. Kumar, S. Mohanty, M. Shahid, A. Sahoo, M. Kaviraj, H. Priya, N.N. Jambhulkar, P.K. Dash, S.D. Mohapatra, P.K. Nayak, Cyanobiont diversity in six *Azolla* spp. and relation to *Azolla*-nutrient profiling, Planta, 249 (2019) 1435–1447.
- [19] Azolla Lam. [family AZOLLACEAE]: JSTOR, 2007 [Updated 19 August 2007]. Available at: https://plants.jstor.org/
- [20] S. Ahmady-Asbchin, A.N. Omran, N. Jafari, Potential of Azolla filiculoides in the removal of Ni and Cu from wastewaters, Afr. J. Biotechnol., 11 (2012) 16158–16164.
- [21] M.A. Zazouli, A.H. Mahvi, S. Dobaradaran, M. Barafrashtehpour, Y. Mahdavi, D. Balarak, Adsorption of fluoride from

aqueous solution by modified *Azolla filiculoides*, Fluoride, 47 (2014) 349–358.

- [22] M.A. Zazouli, Y. Mahdavi, E. Bazrafshan, D. Balarak, Phytodegradation potential of bisphenol A from aqueous solution by *Azolla filiculoides*, J. Environ. Health Sci. Eng., 12 (2014) 66, doi: 10.1186/2052-336X-12-66.
- [23] D. Balarak, E. Bazrafshan, F. Kord Mostafapour, Equilibrium, kinetic studies on the adsorption of Acid Green 3 (AG 3) dye onto *Azolla filiculoides* as adsorbent, J. Am. Chem. Soc., 11 (2016) 1–10.
- [24] A. Ena, P. Carlozzi, B. Pushparaj, R. Paperi, S. Carnevale, A. Sacchi, Ability of the aquatic fern *Azolla* to remove chemical oxygen demand and polyphenols from olive mill wastewater, Grasas y Aceites, 58 (2007) 34–39.
- [25] M. Zhao, J.R. Duncan, R.P. Van Hille, Removal and recovery of zinc from solution and electroplating effluent using *Azolla filiculoides*, Water Res., 33 (1999) 1516–1522.
- [26] M.T. Ganji, M. Khosravi, R. Rakhshaee, Biosorption of Pb, Cd, Cu and Zn from the wastewater by treated *Azolla filiculoides* with H₂O₂/MgCl₂, Int. J. Environ. Sci. Technol., 1 (2005) 265–271.
- [27] R. Rakĥsĥaee, M. Khosravi, M.T. Ganji, Kinetic modeling and thermodynamic study to remove Pb(II), Cd(II), Ni(II) and Zn(II) from aqueous solution using dead and living *Azolla filiculoides*, J. Hazard. Mater., 134 (2006) 120–129.
- [28] S. Prabakaran, T. Mohanraj, A. Arumugam, S. Sudalai, A state-of-the-art review on the environmental benefits and prospects of *Azolla* in biofuel, bioremediation and biofertilizer applications, Ind. Crops Prod., 183 (2022) 114942, doi: 10.1016/j. indcrop.2022.114942.
- [29] R. Sadeghi, R. Zarkami, K. Sabetraftar, P. Van Damme, A review of some ecological factors affecting the growth of *Azolla* spp., Caspian J. Environ. Sci., 11 (2013) 65–76.
- [30] S. Taghilou, M. Peyda, M.M. Mehrasbi, Selection of Wastewater Treatment Process Based on the Analytical Hierarchy Process in Zanjanrood Catchment, 3rd International and 21st National Conference on Environmental Health, Zanjan, Iran, 2019.
- [31] T.A. Lumpkin, Azolla [family AZOLLACEAE]: JSTOR; 2012 [Updated 23 July 2012]. Available at: https://plants.jstor.org/
- [32] P. Brouwer, H. Schluepmann, K.G.J. Nierop, J. Elderson, P.K. Bijl, I. Van Der Meer, W.D. Visser, G.J. Reichart, S. Smeekens, A.V.D. Werf, Growing *Azolla* to produce sustainable protein feed: the effect of differing species and CO₂ concentrations on biomass productivity and chemical composition, J. Sci. Food Agric., 98 (2018) 4759–4768.
- [33] K. Maejima, S. Kitoh, E. Uheda, N. Shiomi, Response of 19 Azolla strains to a high concentration of ammonium ions, Plant Soil, 234 (2001) 247–252.
- [34] A. Golzary, O. Tavakoli, Y. Rezaei, A. Karbassi, Wastewater treatment by *Azolla filiculoides*: a study on color, odor, COD, nitrate, and phosphate removal, Pollution, 4 (2018) 69–76.
- [35] A. Banach, A. Kuźniar, R. Mencfel, A. Wolińska, The study on the cultivable microbiome of the aquatic fern *Azolla filiculoides* L. as new source of beneficial microorganisms, Appl. Sci., 9 (2019) 2143, doi: 10.3390/app9102143.
- [36] M.L. Costa, M.C.R. Santos, F. Carrapiço, A.L. Pereira, Azolla/ Anabaena's behaviour in urban wastewater and artificial media–influence of combined nitrogen, Water Res., 43 (2009) 3743–3750.
- [37] M.J. Pinero-Rodríguez, R. Fernández-Zamudio, I. Gomez-Mestre, C. Díaz-Paniagua, *Ranunculus peltatus* develops an emergent morphotype in response to shading by the invasive *Azolla filiculoides*, Aquat. Bot., 152 (2019) 32–35.
- [38] R.K. Yadav, A. Chatrath, K. Tripathi, M. Gerard, A. Ahmad, V. Mishra, G. Abraham, Salinity tolerance mechanism in the aquatic nitrogen fixing pteridophyte *Azolla*: a review, Symbiosis, 83 (2021) 129–142.
- [39] A. Arora, P.K. Singh, Comparison of biomass productivity and nitrogen fixing potential of *Azolla* spp., Biomass Bioenergy, 24 (2003) 175–178.
- [40] S.J. Varjani, V.K. Srivastava, Green technology and sustainable development of environment, Renewable Resour. J., 3 (2015) 244–249.

- [41] P.S. Parikh, S.K. Mazumder, Capacity of *Azolla pinnata* var. imbricata to absorb heavy metals and fluorides from the wastewater of oil and petroleum refining industry at Vadodara, Int. J. Allied Pract. Res. Rev., 11 (2015) 37–43.
- [42] B. Dhir, P. Sharmila, P.P. Saradhi, Potential of aquatic macrophytes for removing contaminants from the environment, Crit. Rev. Env. Sci. Technol., 39 (2009) 754–781.
- [43] A. Sing, B. Kumar, S. Pabbi, M. Kapoor, Phytoremediation Potential of *Azolla* in Relation to Heavy Metals – A Review, SC, Verma, S.P. Khullar, H.K. Cheema, Eds., Perspect. Pteridophytes, Bishen Singh Mahendra Pal Singh, India, 2008, pp. 487–499.
- [44] A.M. Banach, A. Kuźniar, J. Grządziel, A. Wolińska, Azolla filiculoides L. as a source of metal-tolerant microorganisms, PLoS One, 15 (2020) e0232699, doi: 10.1371/journal.pone.0232699.
- [45] V. Dushenkov, P.B.A.N. Kumar, H. Motto, I. Raskin, Rhizofiltration: the use of plants to remove heavy metals from aqueous streams, Environ. Sci. Technol., 29 (1995) 1239–1245.
- [46] M. Rajkumar, S. Sandhya, M.N.V. Prasad, H. Freitas, Perspectives of plant-associated microbes in heavy metal phytoremediation, Biotechnol. Adv., 30 (2012) 1562–1574.
- [47] A. Banerjee, A. Roychoudhury, Assessing the rhizofiltration potential of three aquatic plants exposed to fluoride and multiple heavy metal polluted water, Vegetos, 35 (2022) 1158–1164.
 [48] M.R.R. Kooh, M.K. Dahri, L.B.L. Lim, L.H. Lim, Batch adsorption
- [48] M.R.R. Kooh, M.K. Dahri, L.B.L. Lim, L.H. Lim, Batch adsorption studies on the removal of Acid Blue 25 from aqueous solution using *Azolla pinnata* and soya bean waste, Arabian J. Sci. Eng., 41 (2016) 2453–2464.
- [49] N. Shafi, A.K. Pandit, A.N. Kamili, B. Mushtaq, Heavy metal accumulation by *Azolla pinnata* of Dal lake ecosystem, India, Dev., 1 (2015) 8–12.
- [50] M. Goala, K.K. Yadav, J. Alam, B. Adelodun, K.S. Choi, M.M.S. Cabral-Pinto, A.A. Hamid, M. Alhoshan, F.A.A. Ali, A.K. Shukla, Phytoremediation of dairy wastewater using *Azolla pinnata*: application of image processing technique for leaflet growth simulation, J. Water Process Eng., 42 (2021) 102152, doi: 10.1016/j.jwpe.2021.102152.
- [51] V. Kumar, P. Kumar, J. Singh, P. Kumar, Potential of water fern (*Azolla pinnata* R.Br.) in phytoremediation of integrated industrial effluent of SIIDCUL, Haridwar, India: removal of physicochemical and heavy metal pollutants, Int. J. Phytorem., 22 (2020) 392–403.
- [52] D. Balarak, E. Bazrafshan, F. Kord Mostafapour, Equilibrium, kinetic studies on the adsorption of Acid Green 3 (AG 3) dye onto *Azolla filiculoides* as adsorbent, Am. Chem. Sci. J., 11 (2016) 1–10.
- [53] P. Carlozzi, G. Padovani, The aquatic fern *Azolla* as a natural plant-factory for ammonia removal from fish-breeding fresh wastewater, Environ. Sci. Pollut. Res., 23 (2016) 8749–8755.
 [54] D.J. Babu, B. Sumalatha, T.C. Venkateswarulu, K.M. Das,
- [54] D.J. Babu, B. Sumalatha, T.C. Venkateswarulu, K.M. Das, V.P. Kodali, Kinetic, equilibrium and thermodynamic studies of biosorption of chromium(VI) from aqueous solutions using *Azolla filiculoides*, J. Pure Appl. Microbiol., 8 (2014) 3107–3116.
- [55] R. Anandha Varun, S. Kalpana, Performance analysis of nutrient removal in pond water using water hyacinth and *Azolla* with papaya stem, Int. Res. J. Eng. Technol., 2 (2015) 444–448.
- [56] A. Arora, S. Saxena, R. Shah, Aquatic Microphyte Azolla for Nutrient Removal from Wastewaters in Constructed Wetlands, Proceedings of International Conference on Energy and Environment, March 19–21, 2009 ISSN:2070-3740, National Institute of Technology, Chandigarh, India, 2009, pp. 185–188.
- [57] N. Shafi, A.K. Pandit, A.N. Kamili, B. Mushtaq, Heavy metal accumulation by *Azolla pinnata* of dal lake ecosystem, India, Dev., 1 (2015) 8–12.
- [58] P.K. Rai, Phytoremediation of Hg and Cd from industrial effluents using an aquatic free floating macrophyte *Azolla pinnata*, Int. J. Phytorem., 10 (2008) 430–439.
- [59] P.K. Rai, B.D. Tripathi, Comparative assessment of Azolla pinnata and Vallisneria spiralis in Hg removal from GB Pant Sagar of Singrauli Industrial region, India, Environ. Monit. Assess., 148 (2009) 75–84.
- [60] P.K. Rai, Microcosm investigation on phytoremediation of Cr using Azolla pinnata, Int. J. Phytorem., 12 (2009) 96–104.

- [61] M.A. Zazoli, D. Belalak, Y. Mahdavi, F. Karimnejad, Application of *Azolla filiculoides* biomass for Acid Blue 15 dye (AB 15) removal from aqueous solutions, J. Basic Res. Med. Sci., 1 (2014) 29–37.
- [62] M.A. Zazouli, D. Balarak, Y. Mahdavi, Effect of *Azolla filiculoides* on removal of Reactive Red 198 in aqueous solution, J. Adv. Environ. Health Res., 1 (2013) 44–50.
- [63] L.J. Umali, J.R. Duncan, J.E. Burgess, Performance of dead *Azolla filiculoides* biomass in biosorption of Au from wastewater, Biotechnol. Lett., 28 (2006) 45–50.
- [64] D. Balarak, R.A. Dianati Tilak, Z. Yousefi, M.A. Zazouli, J. Yazdani, Y. Esfandyari, Phytodegradation potential of phenol from aqueous solution by *Azolla filiculoides*, J. Biorem. Biodegrad., 5 (2014) 66, doi: 10.1186/2052-336X-12-66.
- [65] M.A.Zazouli, A.H.Mahvi, S.Dobaradaran, M.Barafrashtehpour, Y. Mahdavi, D. Balarak, Adsorption of fluoride from aqueous solution by modified *Azolla filiculoides*, Adsorption, 47 (2014) 349–358.
- [66] S. Taghilou, M. Peyda, M.R. Mehrasbi, Modeling of wastewater treatment by *Azolla filiculoides* using response surface methodology, J. Environ. Health Sci. Eng., 19 (2021) 1723–1733.
- [67] P. Jayasundara, Wastewater treatment by Azolla: a review, Diyala Agric. Sci. J., 14 (2022) 40–46.
- [68] P.K. Rai, Phytoremediation of Hg and Cd from industrial effluents using an aquatic free floating macrophyte *Azolla pinnata*, Int. J. Phytorem., 10 (2008) 430–439.
- [69] M.A. Zazouli, Y. Mahdavi, E. Bazrafshan, D. Balarak, Phytodegradation potential of bisphenol A from aqueous solution by *Azolla filiculoides*, J. Environ. Health Sci. Eng., 12 (2014) 66, doi: 10.1186/2052-336X-12-66.
- [70] N.Z. Mamat, S.R.S. Abdullah, H.A. Hasan, N.I. Ismail, S.S.N. Sharuddin, Polishing of treated palm oil mill effluent using *Azolla pinnata*, J. Biochem. Microbiol. Biotechnol., 10 (2022) 40–45.
- [71] L. Polechońska, E. Szczęśniak, A. Klink, Comparative analysis of trace and macro-element bioaccumulation in four freefloating macrophytes in area contaminated by copper smelter, Int. J. Phytorem., 24 (2022) 324–333.
- [72] F.R. El Awady, M.A. Abbas, A.M. Abdelghany, Y.A. El-Amier, Silver modified hydrophytes for heavy metal removal from different water resources, Biointerface Res. Appl. Chem., 11 (2021) 14555–14563.
- [73] L.A. Ndeda, S. Manohar, Bioconcentration factor and translocation ability of heavy metals within different habitats of hydrophytes in Nairobi Dam, Kenya, J. Environ. Sci. Toxicol. Food Technol., 8 (2014) 42–45.
- [74] Q. Hasani, N. Pratiwi, H. Effendi, Y. Wardiatno, J.A.R. Guk Guk, H.W. Maharani, M. Rahman, *Azolla pinnata* as phytoremediation agent of iron (Fe) in ex sand mining waters, J. Nat. Sci., 20 (2021) 1–12.
- [75] I.V. Seregin, A.D. Kozhevnikova, Phytochelatins: sulfurcontaining metal(loid)-chelating ligands in plants, Int. J. Mol. Sci., 24 (2023) 2430, doi: 10.3390/ijms24032430.
- [76] M. Talebi, B.E.S. Tabatabaei, H. Akbarzadeh, Hyperaccumulation of Cu, Zn, Ni, and Cd in *Azolla* species inducing expression of methallothionein and phytochelatin synthase genes, Chemosphere, 230 (2019) 488–497.
- [77] A. Sood, P.L. Uniyal, R. Prasanna, A.S. Ahluwalia, Phytoremediation potential of aquatic macrophyte, *Azolla*, AMBIO, 41 (2012) 122–137.
- [78] L.G. de Araujo, L.C. Vieira, R.L.S. Canevesi, E.A. da Silva, T. Watanabe, R.V. de Padua Ferreira, J.T. Marumo, Biosorption of uranium from aqueous solutions by *Azolla* sp. and *Limnobium laevigatum*, Environ. Sci. Pollut. Res., 29 (2022) 45221–45229.
- [79] R. Bennicelli, Z. Stępniewska, A. Banach, K. Szajnocha, J. Ostrowski, The ability of *Azolla caroliniana* to remove heavy metals (Hg(II), Cr(III), Cr(VI)) from municipal wastewater, Chemosphere, 55 (2004) 141–146.
- [80] M. Hassanzadeh, R. Zarkami, R. Sadeghi, Uptake and accumulation of heavy metals by water body and *Azolla filiculoides* in the Anzali wetland, Appl. Water Sci., 11 (2021) 91, doi: 10.1007/s13201-021-01428-y.

- [81] L.M. Madikizela, Removal of organic pollutants in water using water hyacinth (*Eichhornia crassipes*), J. Environ. Manage., 295 (2021) 113153, doi: 10.1016/j.jenvman.2021.113153.
- [82] I. Maldonado, A.P. Vega Quispe, D. Merma Chacca, F. Zirena Vilca, Optimization of the elimination of antibiotics by *Lemna gibba* and *Azolla filiculoides* using response surface methodology (RSM), Front. Environ. Sci., 10 (2022), doi: 10.3389/fenvs.2022.940971.
- [83] T.J. Al-Musawi, N. Mengelizadeh, M. Taghavi, S. Mohebi, D. Balarak, Activated carbon derived from *Azolla filiculoides* fern: a high-adsorption-capacity adsorbent for residual ampicillin in pharmaceutical wastewater, Biomass Convers. Biorefin., (2021) 1–13, doi: 10.1007/s13399-021-01962-4.
- [84] I. Maldonado, E.G. Moreno Terrazas, F.Z. Vilca, Application of duckweed (*Lemna* sp.) and water fern (*Azolla* sp.) in the removal of pharmaceutical residues in water: state-of-art focus on antibiotics, Sci. Total Environ., 838 (2022) 156565, doi: 10.1016/j.scitotenv.2022.156565.
- [85] A.A. Adelodun, T. Olajire, N.O. Afolabi, A.S. Akinwumiju, E. Akinbobola, U.O. Hassan, Phytoremediation potentials of *Eichhornia crassipes* for nutrients and organic pollutants from textile wastewater, Int. J. Phytorem., 23 (2021) 1333–1341.
- [86] R. Prasad, D. Sharma, K.D. Yadav, H. Ibrahim, Preliminary study on greywater treatment using water hyacinth, Appl. Water Sci., 11 (2021) 1–8.
- [87] A. Sarkar, N. Gogoi, S. Roy, Bisphenol-A incite dosedependent dissimilitude in the growth pattern, physiology, oxidative status, and metabolite profile of *Azolla filiculoides*, Environ. Sci. Pollut. Res. Int., 29 (2022) 91325–91344.
- [88] T. Kösesakal, M. Seyhan, Naphthalene stress responses of the aquatic fern *Azolla filiculoides* Lam. and evaluation of phytoremediation potential, Polycyclic Aromat. Compd., (2022) 1–18, doi: 10.1080/10406638.2022.2126505.
 [89] T. Kösesakal, M. Seyhan, Phenanthrene stress response
- [89] T. Kösesakal, M. Seyhan, Phenanthrene stress response and phytoremediation potential of free-floating fern Azolla filiculoides Lam, Int. J. Phytorem., 25 (2023) 207–220.
- [90] E.Bianchi, A. Biancalani, C. Berardi, A. Antal, D. Fibbi, A. Coppi, L.Lastrucci, N. Bussotti, I. Colzi, L. Renai, C. Scordo, M.D. Bubba, C. Gonnelli, Improving the efficiency of wastewater treatment plants: bio-removal of heavy-metals and pharmaceuticals by *Azolla filiculoides* and *Lemna minuta*, Sci. Total Environ., 746 (2020) 141219, doi: 10.1016/j.scitotenv.2020.141219.
- [91] A. Abd Kadir, S.R.S. Abdullah, B.A. Othman, H.A. Hasan, A.R. Othman, M.F. Imron, N'I. Ismail, S.B. Kurniawan, Dual function of *Lemna minor* and *Azolla pinnata* as phytoremediator for palm oil mill effluent and as feedstock, Chemosphere, 259 (2020) 127468, doi: 10.1016/j.chemosphere.2020.127468.
- [92] National Research Council, Renewable Fuel Standard: Potential Economic and Environmental Effects of U.S. Biofuel Policy, Washington, D.C, 2012, p. 416.
- [93] W. Zhang, A.Y. Elaine, S. Rozelle, J. Yang, S. Msangi, The impact of biofuel growth on agriculture: why is the range of estimates so wide?, Food Policy, 38 (2013) 227–239.

- [94] M. Battaglia, W. Thomason, J.H. Fike, G.K. Evanylo, M.V. Cossel, E. Babur, Y. Iqbal, A.A. Diatta, The broad impacts of corn stover and wheat straw removal for biofuel production on crop productivity, soil health and greenhouse gas emissions: a review, GCB Bioenergy, 13 (2021) 45–57.
 [95] T.G. Ambaye, M. Vaccari, A. Bonilla-Petriciolet, S. Prasad,
- [95] T.G. Ambaye, M. Vaccari, A. Bonilla-Petriciolet, S. Prasad, E.D. Van Hullebusch, S. Rtimi, Emerging technologies for biofuel production: a critical review on recent progress, challenges and perspectives, J. Environ. Manage., 290 (2021) 112627, doi: 10.1016/j.jenvman.2021.112627.
- [96] A. Arora, P. Nandal, A. Chaudhary. Critical evaluation of novel applications of aquatic weed *Azolla* as sustainable feedstock for deriving bioenergy and feed supplement, Environ. Rev., (2022), doi: 10.1139/er-2022-0033.
- [97] H.Z. Hamdan, A.F. Houri, CO₂ sequestration by propagation of the fast-growing *Azolla* spp., Environ. Sci. Pollut. Res., 29 (2022) 16912–16924.
- [98] S. Sathish, S. Supriya, P. Andal, D. Prabu, M. Rajasimman, S. Ansar, S. Rezania, Effective utilization of *Azolla filiculoides* for biodiesel generation using graphene oxide nano catalyst derived from agro-waste, Fuel, 329 (2022) 125412, doi: 10.1016/j.fuel.2022.125412.
- [99] A.F. Miranda, B. Biswas, N. Ramkumar, R. Singh, J. Kumar, A. James, F. Roddick, B. Lal, S. Subudhi, Th. Bhaskar, A. Mouradov. Aquatic plant *Azolla* as the universal feedstock for biofuel production, Biotechnol. Biofuels, 9 (2016) 1–17.
- [100] D. Kannan, W. Christraj, Emission analysis of *Azolla* methyl ester with BaO nano additives for IC engine, Energy Sources Part A, 40 (2018) 1234–1241.
- [101] N. Bose, Production and characterization of biodiesel using Azolla pinnata, Jr. Ind. Pollut. Control, 34 (2018) 1833–1838.
- [102] S.K. Jain, G.S. Gujral, N.K. Jha, P. Vasudevan, Production of biogas from *Azolla pinnata* R. Br and *Lemna minor* L.: effect of heavy metal contamination, Bioresour. Technol., 41 (1992) 273–277.
- [103] A. Mudhoo, S. Kumar, Effects of heavy metals as stress factors on anaerobic digestion processes and biogas production from biomass, Int. J. Environ. Sci. Technol., 10 (2013) 1383–1398.
- [104] C. Paoletti, F. Bocci, G. Lercker, P. Capella, R. Materassi, Lipid composition of *Azolla caroliniana* biomass and its seasonal variation, Phytochemistry, 26 (1987) 1045–1047.
- [105] N.T. Taha Al-Taee, E.S. Mostafa, S.K. AL-Taee, A.A. Abd-Alnafi Al-Aaraji, Impact of *Azolla* on the histopathology of the liver and intestine of the fingerling carp *Cyprinus carpio*, Egypt. J. Aquat. Biol. Fish., 26 (2022) 373–384.
- [106] A.L. Pereira, L.J. Bessa, P.N. Leão, V. Vasconcelos, P.M. da Costa, Bioactivity of *Azolla* aqueous and organic extracts against bacteria and fungi, Symbiosis, 65 (2015) 17–21.